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TITLE: Targeting Tryptophan Catabolism: A Novel Method to Block Triple-Negative Breast Cancer Metastasis

PRINCIPAL INVESTIGATOR: Jennifer Richer

CONTRACTING ORGANIZATION: University of Colorado  
Aurora, CO 80045-2571

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14. ABSTRACT Triple negative breast cancer (TNBC) cells strongly upregulate two related pathways, the kynurenine pathway (KP) in forced suspension culture. The KP is the principal route of tryptophan catabolism. Interestingly, the intermediate tryptophan metabolite kynurenine (Kyn) was recently identified as an endogenous ligand for AhR, a transcription factor that was also upregulated in suspension. Kyn activation of AhR promotes motility of glioma cells, and increased AhR expression and activity was found to be a characteristic of human and mouse mammary tumors. AhR is also in many immune cell types and its activation decreases T-cell activity leading to tumor immune escape. Since the rate limiting enzyme TDO2 increases in TNBC, the goal of our proposal is to determine if we can target this pathway that helps TNBC metastasize. Our hypothesis is that upregulation of kynerinine by TNBC facilitates survival in transit to metastatic sites and immune suppression and thereby mediates the highly metastatic nature of this subtype. We now have one published work and work in progress suggests that this hypothesis holds true.					
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## 1. INTRODUCTION:

- KEYWORDS:** Triple negative breast cancer (TNBC), kynurenine, tryptophan metabolism, anoikis resistance, metastasis
- ACCOMPLISHMENTS:** **What were the major goals of the project?** Our overall goals were to test the hypotheses that the ability to upregulate kynurenine via the enzyme TDO2 supports the highly metastatic nature of TNBC by two mechanisms since the receptor for kynurenine (KYN), AhR, is expressed in both TNBC cells and effector T cells, such that Kyn binding to AhR elicits both autocrine effects helping the tumor cells survive in transit as well as paracrine immune-suppressive effects.

**The major goals as stated in the approved SOW are below along with completion status.**

**Aim 1: Determine if inhibition of TDO2 affects TNBC cell migration, invasion, and anchorage independent growth and if these effects are mediated by kynurenine activation of AhR. (Months 1-12)**

**Task1. Confirm that production of Kynurenine increases in TNBC cells in suspension and that this is mediated by TDO2. Compare 3 TNBC lines (BT549, SUM-159 MDA-231) to 3 luminal lines (MCF7, ZR-75.1, T47D). (Months 1-2) Complete**

**The figures referred to herein in Aims 1 and 2 (the first part of the SOW and progress report) refer to our published paper (D'Amato and Rogers et al Cancer Research, 2015), which is attached in the appendix.** To confirm our gene array data, we performed qRT-PCR for *TDO2* and *KYNU* in multiple breast cancer cell lines, including both luminal (ER+; MCF7 and T47D) and TNBC (ER-; MDA-231, BT549, and SUM159) lines, after 24hrs in suspension (**Figure 1B and C**). In all three TNBC lines tested, *TDO2* and *KYNU* were significantly increased in suspension compared to attached culture. In the two ER+ breast cancer cell lines tested, expression of these genes trended slightly higher in suspension but this change was not significant. Western blot analysis of whole cell extracts also demonstrated an increase in TDO2 and KYNU protein in TNBC cell lines (MDA-231, BT549, and SUM159) grown in suspension for 24hrs (Figure 1D and Supplemental Figure 3A). The increase in TDO2 protein was confirmed by IHC in BT549 cells grown in suspension for 48hrs compared to cells grown in the attached condition (**Figure 1E**). Global metabolomic profiling of intracellular and secreted metabolites from BT549 cells grown in standard attached conditions or in forced suspension for 24 hours was also performed. Two intermediate products of the kynurenine pathway, Kyn and formylkynurenine, were the intracellular metabolites with the highest fold-change increase in suspension. Among secreted metabolites, kynurenine had the third-highest fold-change increase (Supplemental Figure 2). Together with the gene expression data, this demonstrates that the kynurenine pathway is strongly upregulated in TNBC cells upon loss of attachment. Using HPLC to verify the metabolomic profiling data, we found that secreted Kyn levels were more than two-fold higher in conditioned media from BT549 cells in forced suspension for 48hrs than in media from the same number of cells in the attached condition (Figure 1F). Furthermore, addition of the TDO2-specific inhibitor 680C91 to cells in suspension completely prevented the increase in secreted Kyn, demonstrating that increased secretion of Kyn in TNBC cells in suspension is dependent on TDO2 activity (Figure 1F).

**Task 2. Determine if TDO2 or AhR knockdown or inhibition decreases migration (Months 1-4) Complete** Pharmacological inhibition of TDO2 and AhR each significantly reduced migration of MDA-MB-231 and BT549 cells in a scratch wound assay, and again the combination was more effective than either inhibitor alone (Figure 4B). Knockdown of either TDO2 or AhR recapitulated this effect, significantly diminishing migration in the scratch wound assay in both BT549 and MDA-231 cells (**Figure 4C and 4D**).

**Task 3. Determine if TDO2 or AhR knockdown or inhibition decreases invasion (Months 5-8) Complete**

To test whether the increased TDO2 and AhR expression observed in cells grown in suspension affects invasive capacity, SUM159PT cells were grown for 48hrs either in the attached condition, or in forced suspension culture with or without addition of 10μM of the TDO2 inhibitor 680C91 or the AhR inhibitor CH-223191. After

24hrs, 25,000 viable cells were plated in a Matrigel-coated trans-well invasion chamber with continuous treatment. Cells grown in suspension for 24hrs were significantly more invasive than control cells grown in the attached condition (**Figure 4E**). The addition of the TDO2 inhibitor 680C91 during suspension culture greatly decreased the invasive capacity of viable cells. While the cells treated with the AhR inhibitor showed decreased invasion, this effect was not statistically significant (**Figure 4E**).

**Task 4. Test if inhibition or knockdown of TDO2 or AhR decrease anchorage-independent growth of TNBC cells in vitro (Months 9-12) Completed**

Since TDO2 and AhR were upregulated by TNBC cells in suspension, we next tested their functional importance by assessing whether pharmacological inhibition or genetic knockdown could reduce anchorage-independent growth in soft agar or increase sensitivity of TNBC cells to anoikis in forced suspension culture. BT549 and SUM159PT cells were pre-treated in the attached condition with vehicle, the TDO2 inhibitor 680C91, or the AhR antagonist CH-223191 for 24hrs. Then an equal number of cells were plated in soft agar in the continued presence of treatment. Both 680C91 and CH-223191 significantly decreased anchorage-independent growth of BT549 and SUM159PT cells in soft agar (**Figure 3A**). To test the effect of Kyn on anoikis resistance, we treated cells in forced suspension culture for 48hrs with Kyn, and found that this significantly decreased apoptosis as measured by cleaved caspase activity compared to vehicle control treatment (**Figure 3B**). We then performed knockdown of TDO2 and AhR using two shRNA constructs each, and decreased protein expression was confirmed by western blot (**Figure 3C**). Knockdown of either TDO2 or AhR also significantly decreased growth of BT549 cells in soft agar (**Figure 3D**). Knockdown of either TDO2 or AhR also significantly increased apoptosis in BT549 and MDA-MB-231 cells grown in suspension for 48hrs (**Figure 3E**), demonstrating that TDO2 and AhR promote survival in anchorage-independent conditions. We have yet to determine if overexpression of TDO2 increases anchorage independent in vitro, but we have acquired a TDO2 overexpression vector that can be stably transduced into breast cancer cell lines (TNBC, luminal (ER+), immortalized normal mammary epithelial cells like the MCF-10A) and selected using puromycin.

**Aim 2: Test whether inhibition of TDO2 and AhR affects the ability of TNBC to survive in the bloodstream following tail vein injection or to metastasize in vivo from the orthotopic site. (Months 12-24) we are a bit ahead of schedule because we wanted to get the results published in a good journal, so needed an in vivo experiment.**

**Task 1. Determine if TDO2 activity is critical for short term survival of TNBC cells in the vasculature (testing anoikis resistance in vivo) (Months 13-16) (underway)**

Still in progress.

**Task. 2. Determine if pharmacological inhibition of TDO2 decreases metastasis following tail vein injection (Months 17-20) (completed)**

Because we needed this in vivo test to get the work published, we went ahead and did this experiment. To test the contribution of TDO2 activity to the metastatic capacity of TNBC cells in vivo, we grew luciferase-expressing MDA-MB-231 cells in forced suspension conditions for 48hrs in the presence of either vehicle control or the TDO2 inhibitor 680C91. 250,000 viable cells, as determined by trypan blue staining, were then injected into the tail vein of NOD/SCID mice and luminescence was monitored over time. Seven days after injection, mice that received vehicle-treated cells had significantly higher luminescence, and this statistically significant difference was maintained throughout the experiment (**Figure 6A-B**). At the conclusion of the experiment at Day 28 post-injection, lung luminescence was measured ex-vivo, and lungs from mice that received vehicle-treated cells had significantly higher luminescence compared to mice that received cells treated with the TDO2 inhibitor (**Figure 6C**). A significant decrease in the number of metastatic nodules in the lungs from mice receiving cells treated with 680C91 was also observed by H&E (**Figure 6D**). Together, these data demonstrate that TDO2 inhibition decreases the ability of TNBC cells to successfully metastasize following tail vein injection in vivo.

**Aim 3: Identify the role of kyn paracrine action in tumor-induced immune modulation via AhR-expressing effector T cells. (Months 1-36) (60% complete)**

1. Determine whether conditioned media from TNBC in suspension with or without TDO inhibition alters the number and function of effector T cells via the Kyn-AhR pathway (Months 1-15) (Complete)

a. **Isolate CD8+ and CD4+ T cells from normal donors and breast cancer patients.** CD8+ and CD4+ T-cells were both successfully isolated from the blood of healthy donors (no cancer) under the IRB-approved protocol lead by Dr. Virginia Borges using positive selection kits (Dynabeads, Invitrogen) (A pure population of CD3+ CD8+ T-cells from the blood of a healthy volunteer donors is shown in **Figure A.**). The isolation of these T-cells from the blood of breast cancer patients is currently in progress.

b. **Activate T cells by coculture with  $\alpha$ CD3 and  $\alpha$ CD28 in the presence of conditioned media of TNBC cells (SUM-159 and BT549) grown in either attached condition, suspended, or suspended + 680C91.** T-cells were successfully activated over the course of 5 days using soluble anti-CD28 (1 ug/ml) and plate-bound anti-CD3 (0.5 ug/ml). The concentration of anti-CD3 and the number of T-cells plated were optimized to ensure good activation without over-stimulating the T-cells (**Figure B**).

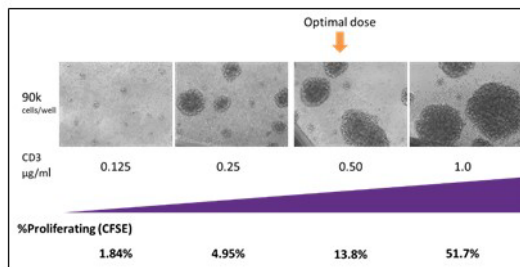


Figure B: Optimization of T-cell activation from the blood of normal human donors. T-cells were isolated as described above, then activated over the course of 5 days using plate-bound anti-CD3 (concentrations shown) and 1 ug/ml soluble anti-CD28. Cell number was also optimized (90000 cells per well) to determine the best parameters for in vitro human T-cell activation.

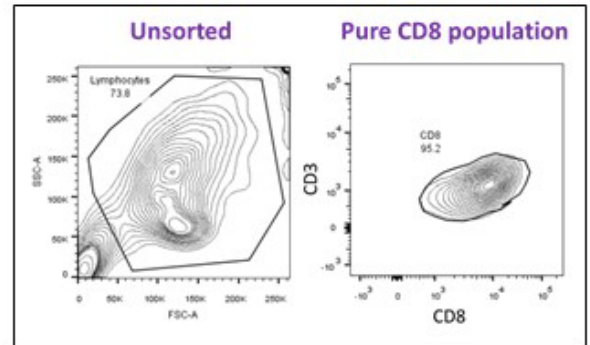


Figure A: isolation of CD8+ T-cells from the blood of normal donors. Lymphocytes were isolated using Ficoll separation, and then CD8+ T-cells were selected using a positive selection kit (Dynabeads, Invitrogen). Cells were then stained for CD3 and CD8, and a highly pure (95.2% pure) population of CD8+ T-cells was identified.

c. **Analyze T cell subsets by flow cytometry, and test proliferative capacity by CFSE, T cell viability using a fixable viability stain, and the generation of Treg cells (CD4+,FoxP3+,CD25+).** After the 5-day activation in conditioned media from TNBC cells that were grown in attached, suspended, or suspended + 680C91 conditions, T-cells were analyzed using flow

cytometry. The proliferative capacity of the T-cells was measure by CFSE dilution (%diluted), and T-cell death was measured by incorporation of a fixable viability dye (eBioscience). The results of these experiments, repeated in 10-12 healthy donors (cancer donor blood in progress), is in **Figure C**.

d. We found that conditioned media from attached BT549 cells increases CD8 T-cell death and decreases CD8-T-cell proliferation, indicating suppression of these T-cells. Our preliminary data also suggest that 680C91 could contribute to lessening the effect of suspended conditioned media on the CD8 T-

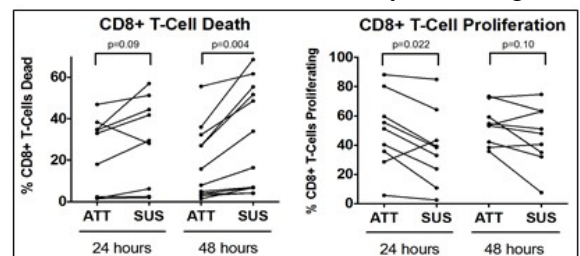


Figure C: conditioned media from suspended BT549 cells increases cell death and decreases proliferation in primary human CD8+ T-cells. BT549 cells were plated in attached or suspended conditions for either 24 or 48 hours. Conditioned media was then collected, filtered, and human CD8 T-cells (isolated as described) were plated in these conditioned media over the course of a 5-day activation (optimized in Figure B). Cell death was measured by incorporation of a fixable viability dye (eBioscience) and proliferation was measured by dilution of the cell-permeable dye CFSE (% diluting) using flow cytometry. Paired T-test, n=10-12 donors.

cells (**Figure D**). The impact of conditioned media on T-regs is still being investigated, but we were able to detect an increase in CD4<sup>+</sup>,CD25<sup>+</sup>,Foxp3<sup>+</sup> cells in the presence of purified kynurenine (from an isolated population of CD3<sup>+</sup> CD4<sup>+</sup> T-cells from healthy donor blood, **Figure E**).

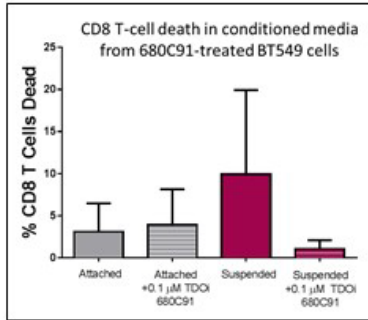


Figure D: CD8 T-cells were plated in conditioned media from BT549 cells that had been in attached or suspended conditions for 48 hours with or without 680C91 (0.1 μM). Cell viability was then measured using a fixable viability dye and flow cytometry as described above. Preliminary data, n=2 donors.

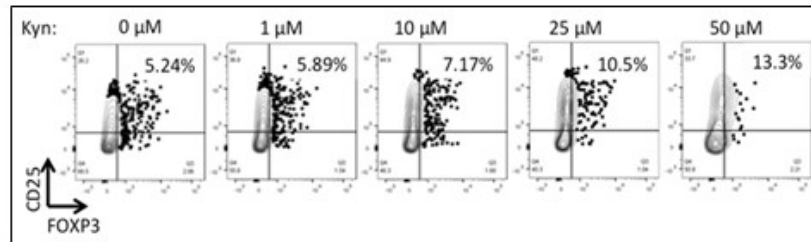


Figure E: CD4<sup>+</sup> T-cells were isolated as described and then treated with purified kynurenine at the indicated doses over the course of a 5-day CD3/CD28 activation. Using flow cytometry, CD3<sup>+</sup>/CD4<sup>+</sup> T-cells were analyzed for their expression of the T-reg markers CD25 and FoxP3 (an intracellular stain).

- e. **Test for functional Treg induction by coculture of stimulated CD8<sup>+</sup> T cells from normal donors with CD4<sup>+</sup> FoxP3<sup>+</sup>CD25<sup>+</sup> Treg cells generated in the presence of conditioned media from TNBC cells grown in either attached condition, suspended, or suspended + 680C91.** The T-reg functionality assays are being optimized with the help of Dr. Jill Slansky's laboratory.
- f. **Test in vitro for cytotoxicity function of CD8<sup>+</sup> T-cells treated with conditioned media from TNBC cells grown in attached, suspended, or suspended + 680C91 (or CD8<sup>+</sup> T-cells treated with Kyn) in killing tumor cells.** This in vitro killing assay is also still being optimized with the help of Dr. Slansky's laboratory.
2. Analyze the cytokine production of T cells stimulated with and without the TNBC-conditioned media and determine if similar cytokines are present in women with metastatic TNBC, including the AhR expression in their T cells in comparison to healthy donors (Months 5-15). (40% complete)
  - a. **Isolate CD8<sup>+</sup> and CD4<sup>+</sup> T cells from normal donors and breast cancer patients.** See Aim 3.1.A and Figure A.
  - b. **Activate T cells by coculture with αCD3 and αCD28 in the presence of conditioned media of TNBC cells grown in either attached condition, suspended, or suspended + 680C91.** See Aim 3.1.B and Figures C and D.
  - c. **Analyze cytokines produced by T cells using Proinflammatory Panel 1 multiplex detection kit (MesoScale Discovery) and by using intracellular stains for interferon-gamma and granzyme-B followed by flow cytometry.** We are still optimizing the Mesoscale multiplex detection kit for this purpose, but we have used intracellular stains for interferon-gamma and granzyme B and analyzed these stains using flow cytometry. We found that conditioned media from suspended TNBC cells (BT549) does reduce interferon gamma production by CD8 T-cells. The impact of these conditioned media on CD4 T-cells is still being investigated (Figure F).



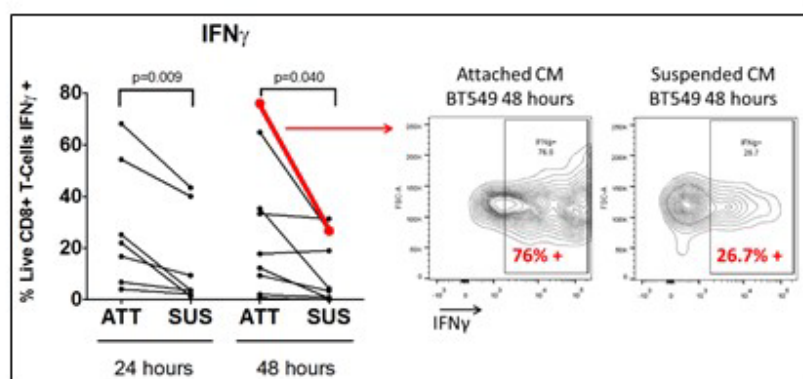


Figure F: CD8 T-cells were isolated as described and cultured in conditioned media from attached or suspended BT549 cells as described over the course of a 5-day CD3/CD28 activation. Interferon gamma positivity was measured using an intracellular stain analyzed via flow cytometry. N=8 healthy human donors, paired t-test.

- d. **Perform western blotting and qRT-PCR to assess levels of AhR and AhR target genes such as CYP1A1 and CYP1B1 in T cells culture in different conditioned medias.** These experiment are in progress.
- e. **Perform western blotting using nuclear/cytosolic fractionation to assess changes in nuclear localization of AhR in T cells cultured in different conditioned medias.** These experiments are in progress.

### What opportunities for training and professional development has the project provided?

Two graduate students, Thomas Rogers and Lisa Greene have been working on this project. The abstracts for talks they have presented are listed below in section 6 on products and shown in the appendix.

Thomas also gave an invited oral presentation at the University of Colorado Cancer Center retreat entitled “Metabolism and Cancer” hosted by the Molecular Oncology Program.

Thomas was awarded the 2015 Denver School of Science and Technology-Anschutz Medical Campus Teaching Fellowship where he designed and taught a health science elective for high school students to prepare them for internships in the health sciences. Additionally, he participated in a pilot workshop on “Learning How to Teach” supported by the Broadening Experiences in Scientific Training (BEST) Program awarded to the University of Colorado by the NIH. This workshop, led by a group of experienced teachers from our institution, is aimed at helping predoctoral trainees prepare to teach at the college level. Furthermore, Thomas taught in CLSC 7500 (*Practical Application of Molecular and Cell Biology Techniques for the Clinical Investigator*) during the month of July 2015 designed to train clinical investigators laboratory techniques that they could apply to their research interests.

Thomas Rogers obtained the following training grant and was the sole student selected from a competitive pool at our institution to submit a new grant mechanism, the NCI Predoctoral to Postdoctoral Fellow Transition Award (F99/K00)

**Current training grant:** 1F31CA203394-01

Title: *Targeting a Kynurenine-Driven Autocrine Loop to Block Triple-Negative Breast Cancer Metastasis*



Amount: \$32,921/year

Dates: 03/07/2016-3/07/2019

**Pending Research Support:**

1F99CA212230-01

Title: *Targeting a Kynurenine-Driven Autocrine Loop to Block Triple-Negative Breast Cancer Metastasis*

Lisa has also submitted a predoctoral NRSA on her project last month that will be reviewed June 27<sup>th</sup>.

**How were the results disseminated to communities of interest?** Article on the beginnings of this work in [http://www.eurekalert.org/pub\\_releases/2014-12/uocd-ptd121014.php](http://www.eurekalert.org/pub_releases/2014-12/uocd-ptd121014.php)

**What do you plan to do during the next reporting period to accomplish the goals?**

Determine if treatment with a small molecule inhibitor of TDO2 decreases metastasis of TNBC cells from an orthotopic xenograft model (**Months 21-24**). As mentioned in our SOW, we also planned on treating the mice systemically with 680C91 in their drinking water and then observing if TNBC metastasis was affected. However, although it has been used successfully in rodent models before, the TDO2 inhibitor 680C91 has been previously shown to have poor stability *in vivo*. To circumvent this issue, we have acquired a 2<sup>nd</sup> generation TDO2 inhibitor (LM10) from the Ludwig Institute for Cancer Research in Belgium through a materials transfer agreement (MTA). Since we are just going into month 13 we will first be characterizing this compound *in vitro* before treating mice.

We will also be conducting Task Aim 3 to Determine if metastatic potential of a syngeneic mouse breast cancer model can be altered by TDO inhibition and whether there is alteration of the tumor immune microenvironment with decreased infiltration and/or function of the tumor-infiltrating lymphocytes (**Months 15-36**).

**4. IMPACT:**

- **What was the impact on the development of the principal discipline(s) of the project? "**
- This research is strongly suggesting that TDO2 is likely the primary enzyme that catabolizes tryptophan that should be targeted in breast cancer and that by targeting this enzyme we may be able to reduce breast cancer metastasis by decreasing the tumor cell's ability of the cells to survive in suspension, but also reverse their ability to suppress the immune system.
- **What was the impact on other disciplines?** Nothing to Report.
- **What was the impact on technology transfer?**
  - This project is likely to make an on commercial technology because it shows evidence that TDO2 is likely the primary enzyme that catabolizes tryptophan that should be targeted in breast cancer. The current drugs that are in clinical trials target IDO, not TDO2.
- **What was the impact on society beyond science and technology?** Gave two trainees great opportunities to present their work and further their career development.

**5. CHANGES/PROBLEMS: e:**

- **Changes in approach and reasons for change** *None*
- **Actual or anticipated problems or delays and actions** *None*
- **Changes that had a significant impact on expenditures** *None*
- **Significant changes in use or care of human subjects, vertebrate animals, biohazards, and/or select agents**
  - *Also specify the applicable Institutional Review Board/Institutional Animal Care and Use Committee approval dates.*
- **Significant changes in use or care of human subjects.** *None*
- **Significant changes in use or care of vertebrate animals.** *None*
- **Significant changes in use of biohazards and/or select agents.** *None*

## 6. PRODUCTS:

- **Publications, conference papers, and presentations**

### **Journal publications:**

1. D'Amato NC\*, **Rogers TJ\***, Gordon MA, Greene LI, Cochrane DR, Spoelstra NS, Nemkov TG, D'Alessandro A, Hansen KC, Richer JK. A TDO2-AhR Signaling Axis Facilitates Anoikis Resistance and Metastasis in Triple-Negative Breast Cancer. *Cancer Res.* 2015 Nov 1;75(21):4651-64. PubMed PMID: [26363006](#); PubMed Central PMCID: [PMC4631670](#).

Federal support was acknowledged

### **Conference presentations and posters:**

4<sup>th</sup> year Cancer Biology Graduate Program Doctoral Candidate **Thomas Rogers** attended the 2015 **International Society for Tryptophan Research Conference in September 2015**, at Van Andel Research Institute in Grand Rapids, Michigan and was invited to give a Late-Breaking Oral Presentation entitled ***“Targeting a Kynurenine-Driven Autocrine Loop to Block Triple-Negative Breast Cancer Metastasis”***. His presentation was given in front of many experts in the field of tryptophan catabolism across a number of different disease types and he was awarded the Best Late-Breaking Oral Presentation at this conference and formed many good interactions/collaborations.

Thomas also attended the 2016 Keystone Symposium- Nuclear Receptors meeting and gave a poster on ***“Endogenous activation of the aryl hydrocarbon receptor promotes triple-negative breast cancer metastasis”***.

Cancer Biology Graduate Program Doctoral Candidate **Lisa Greene** gave a short talk **“Suppression of CD8 T-cells by tryptophan catabolism in triple-negative breast cancer”** Lisa I. Greene, Tullia C. Bruno, Thomas J. Rogers, Jill E. Slansky, Virginia F. Borges, Jennifer K. Richer at the **Keystone Symposium on Nuclear Receptors in Jan 2016**.

- **Website(s) or other Internet site(s)**  
None at this time.
- **Technologies or techniques**  
None at this time.
- **Inventions, patent applications, and/or licenses**  
None at this time.
- **Other Products**  
*Identify any other reportable outcomes that were developed under this project. Examples include:*
  - data or databases: none at this time
  - biospecimen collections: FFPE blocks of lungs with metastases of MDA-MB-231 xenografts of tumors pretreated with or without TDO2 inhibitor
  - research material: knowledge of PDX that express TDO2 or AhR.

## 7. PARTICIPANTS & OTHER COLLABORATING ORGANIZATIONS

- **What individuals have worked on the project?**

4/2015-3/2016	Cal Months
RICHER, JENNIFER	1.12
SLANSKY, JILL	0.6
D'ALESSANDRO, ANGELO	1.2
BORAKOVE, MICHELLE	1.8
SPOELSTRA, NICOLE	2.76
CARSON BUTTERFIELD, KIE	6
BRUNO, TULLIA	0.4
GREENE, LISA	8.67
ROGERS, THOMAS	6
MOORE, BRANDON	0.4

o

Researcher Identifier CAGE code is 0P6C1 for all

- o **Has there been a change in the active other support of the PD/PI(s) or senior/key personnel since the last reporting period?**
  - No
- o **What other organizations were involved as partners?**
  - No

#### 8. SPECIAL REPORTING REQUIREMENTS N/A

#### 9. APPENDIX

# A TDO2-AhR Signaling Axis Facilitates Anoikis Resistance and Metastasis in Triple-Negative Breast Cancer

Nicholas C. D'Amato<sup>1</sup>, Thomas J. Rogers<sup>1</sup>, Michael A. Gordon<sup>1</sup>, Lisa I. Greene<sup>1</sup>, Dawn R. Cochrane<sup>1</sup>, Nicole S. Spoelstra<sup>1</sup>, Travis G. Nemkov<sup>2</sup>, Angelo D'Alessandro<sup>2</sup>, Kirk C. Hansen<sup>2</sup>, and Jennifer K. Richer<sup>1</sup>

## Abstract

The ability of a cancer cell to develop resistance to anoikis, a programmed cell death process triggered by substratum detachment, is a critical step in the metastatic cascade. Triple-negative breast cancers (TNBC) exhibit higher rates of metastasis after diagnosis, relative to estrogen-positive breast cancers, but while TNBC cells are relatively more resistant to anoikis, the mechanisms involved are unclear. Through gene expression and metabolomic profiling of TNBC cells in forced suspension culture, we identified a molecular pathway critical for anchorage-independent cell survival. TNBC cells in suspension upregulated multiple genes in the kynurenine pathway of tryptophan catabolism, including the enzyme tryptophan 2,3-dioxygenase (TDO2), in an NF- $\kappa$ B-dependent manner. Kynurenine production mediated by TDO2 in TNBC cells was sufficient to activate aryl hydrocarbon

receptor (AhR), an endogenous kynurenine receptor. Notably, pharmacologic inhibition or genetic attenuation of TDO2 or AhR increased cellular sensitivity to anoikis, and also reduced proliferation, migration, and invasion of TNBC cells. *In vivo*, TDO2 inhibitor-treated TNBC cells inhibited colonization of the lung, suggesting that TDO2 enhanced metastatic capacity. In clinical specimens of TNBC, elevated expression of TDO2 was associated with increased disease grade, estrogen receptor-negative status, and shorter overall survival. Our results define an NF- $\kappa$ B-regulated signaling axis that promotes anoikis resistance, suggest functional connections with inflammatory modulation by the kynurenine pathway, and highlight TDO2 as an attractive target for treatment of this aggressive breast cancer subtype. *Cancer Res*; 75(21); 4651–64. ©2015 AACR.

## Introduction

The vast majority of breast cancer deaths are caused by complications from metastases (1–3). A high rate of metastasis is characteristic of triple-negative breast cancers (TNBC), which lack expression of estrogen receptor (ER), progesterone receptor (PR), and HER2 amplification, and thus do not respond to current endocrine therapies or HER2-targeted therapies (4). Due in part to the absence of effective targeted therapies, but likely also to inherent properties of this subtype, patients with metastatic TNBC have a poor prognosis with a median survival of 13 months (5). Thus, identification of new targeted therapies that inhibit or slow metastasis is of critical importance to improve the prognosis of women with TNBC.

Metastasis is a complex process with multiple steps, including detachment of cancer cells from the primary tumor, invasion through local tissue, intravasation into the vasculature and lymphatics, survival while in transit, extravasation, and colonization of secondary sites (6). Normal epithelial cells are programmed to undergo apoptosis if they become detached from the basement membrane, a process termed anoikis (7). Anoikis resistance is thought to facilitate survival of tumor cells that detach from the primary tumor and thereby facilitate metastasis (8, 9). Multiple mechanisms of anoikis resistance have been identified, including deregulation of integrin expression and aberrant activation of prosurvival pathways (as reviewed in ref. 9), as well as altered metabolism (10, 11). However, the present study is the first to globally profile gene expression alterations in forced suspension culture with the goal of identifying targetable pathways important for survival in suspension. This objective screening approach identified multiple components of the kynurenine pathway of tryptophan catabolism and the aryl hydrocarbon receptor (AhR), which is activated by kynurenine (12), as being upregulated by TNBC in suspension culture.

Altered tryptophan metabolism and increased secretion of tryptophan metabolites by solid tumors, including in breast cancer, have long been recognized (13, 14). The essential amino acid tryptophan is required for protein synthesis and is a precursor for the formation of multiple signaling molecules including serotonin (15). The majority of tryptophan catabolism occurs via the kynurenine pathway, leading to synthesis of NAD<sup>+</sup> along with intermediate products, including quinolinic acid and

<sup>1</sup>Department of Pathology, University of Colorado Anschutz Medical Campus, Aurora, Colorado. <sup>2</sup>Department of Biochemistry and Molecular Genetics, University of Colorado Anschutz Medical Campus, Aurora, Colorado.

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N.C. D'Amato and T.J. Rogers contributed equally to this article.

**Corresponding Author:** Jennifer K. Richer, University of Colorado at Denver, RCI North P18-5127 Mail Stop 8104, 12800 E. 19th Ave, Aurora, CO 80045. Phone: 303-724-3735; Fax: 303-724-3712; E-mail: [Jennifer.Richer@ucdenver.edu](mailto:Jennifer.Richer@ucdenver.edu)

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kynurenine (Kyn; ref. 16). The first step of the kynurenine pathway can be catalyzed by indoleamine 2,3-dioxygenase 1 (IDO1), IDO2, or tryptophan 2,3-dioxygenase (TDO2; refs. 17–19). IDO1 is expressed in tissues throughout the body, whereas TDO2 is predominantly expressed in the liver (20). Much research on the kynurenine pathway in cancer has focused on the ability of Kyn to decrease immune surveillance (as reviewed in ref. 21), but in glioma, Kyn, acting in both a paracrine and an autocrine fashion, suppressed antitumor immune responses and promoted tumor cell survival and motility. Kyn was demonstrated to serve as an endogenous ligand to activate AhR in both immune cells and the tumor cells themselves (12).

AhR is a member of the basic-helix-loop-helix (bHLH) Per-ARNT-Sim (PAS) superfamily of transcription factors best known for its activation by xenobiotics such as polycyclic aromatic hydrocarbons (PAH; refs. 22, 23). PAH binding leads to transcription of AhR target genes, including drug-metabolizing enzymes such as *CYP1A1* and *CYP1B1* (24). Increased expression of AhR and its target genes has been found in several cancer types, including lung, cervical, ovarian, and breast (16). AhR is required for normal mammary gland development (25, 26), and AhR overexpression increased migration, invasion, and proliferation of immortalized mammary epithelial cells (27). AhR knockdown in MDA-MB-231 TNBC cells decreased expression of genes involved in these same processes (28). Interestingly, increased AhR activity in mouse hepatoma cells grown in suspension was observed nearly 20 years ago (29); however, neither the mechanism of activation nor the functional significance was tested.

Using global expression analysis and metabolomic profiling of TNBC cells in forced suspension, we identify a novel TDO2-AhR signaling axis, mechanistically dependent on NF- $\kappa$ B, which promotes anoikis resistance, as well as migratory and invasive capacity. Indeed, we found that pharmacologic inhibition or knockdown of TDO2 or AhR decreased anchorage-independent growth and invasive capacity *in vitro* and TDO2 inhibition decreased lung metastasis in a TNBC preclinical model. Collectively, these data and the fact that *TDO2* expression confers a shorter overall survival in breast cancer patients suggest that TDO2 inhibition may be a rational targeted therapy to reduce TNBC metastasis and resultant mortality.

## Materials and Methods

### Cell culture and treatments

All cell lines were authenticated by short tandem repeat analysis and tested negative for mycoplasma in July of 2014. SUM159PT cells were purchased from the University of Colorado Cancer Center Tissue Culture Core in August of 2013 and were grown in Ham's F-12 with 5% FBS, penicillin/streptomycin, hydrocortisone, insulin, HEPES, and L-glutamine supplementation. MDA-MB-231 (MDA-231) cells were purchased from the ATCC in August of 2008 and were grown in minimum essential media with 5% FBS, penicillin/streptomycin, HEPES, L-glutamine, non-essential amino acids, and insulin supplementation. BT549 cells, purchased from the ATCC in 2008, were grown in RPMI Medium 1640 with 10% FBS, penicillin/streptomycin, and insulin. MCF7 and T47D cells were purchased from the ATCC and were grown in DMEM with 10% FBS and 2 mmol/L L-glutamine.

### Forced suspension culture

Poly-2-hydroxyethyl methacrylate (poly-HEMA, from Sigma-Aldrich) was reconstituted in 95% ethanol to 12 mg/mL. Ethanol

was allowed to evaporate overnight, and plates were washed with PBS prior to use.

### Kynurenine high performance liquid chromatography

Experimental samples were generated by plating 500,000 cells in 2 mL culture media in a 6-well plate in triplicate. After 48 hours, media were collected and protein was precipitated using trichloroacetic acid at a final concentration of 7%, samples were centrifuged, and the supernatant was analyzed on an Agilent 1260 high performance liquid chromatography (HPLC) with a Kinetex BiPhenyl column (particle size 2.7  $\mu$ m, 4 mm internal diameter by 150 mm; Phenomenex, catalog no. 00F-4622-E0). Kynurenine was eluted using a binary gradient consisting of 0.2% formic acid in water (mobile phase A) and 0.2% formic acid in acetonitrile (mobile phase B). The separation gradient was 5% to 60% B over 6 minutes at a flow rate of 0.75 mL/min, and the kynurenine peak was monitored at 360 nm. To quantify kynurenine in the samples, peak areas were compared with a serial dilution of L-kynurenine (Sigma; catalog no. K8625) in tissue culture media, which was prepared with the same precipitation method as the samples. In brief, 100  $\mu$ L of standard or supernatant was injected onto the system, and peak area was calculated using Agilent ChemStation software.

### Cellular assays and reagents

Cells were treated with 680C91 and CH223191 (TOCRIS Bioscience) prepared in dimethyl sulfoxide (DMSO). Alpha-naphthoflavone (Sigma-Aldrich) was diluted in methanol:ethyl acetate (3:1), and 1-D,L-methyl-tryptophan and crystalline L-kynurenine (Sigma-Aldrich) were prepared in 0.25N HCl. PS 1145 (TOCRIS Bioscience) was prepared in DMSO. IL1 $\beta$  and TNF $\alpha$  were obtained from eBioscience.

### Cellular fractionation

Cellular fractionation was performed using the NE-PER Nuclear and Cytoplasmic Extraction Kit (Pierce Biotechnology) as per the manufacturer's instructions.

### Quantitative RT-PCR

Total RNA was isolated using the RNeasy Mini Plus Kit (QIAGEN) according to the manufacturer's instructions. cDNA was synthesized with the Applied Biosystems reverse transcription Kit. qRT-PCR was performed in an ABI 7600 FAST thermal cycler using Absolute Blue qPCR SYBR Green Low ROX Mix (Thermo Scientific). All fold-change data were normalized to  $\beta$ -actin. All experiments were performed in biologic triplicate. For primer sequences, see Supplementary Methods.

### Immunoblotting

Whole-cell protein extracts (50  $\mu$ g) were denatured, separated on SDS-PAGE gels, and transferred to polyvinylidene fluoride membranes. After blocking in 3% BSA in Tris-buffered saline-Tween, membranes were probed overnight at 4°C. Primary antibodies used include AhR (#13790; 1:1,000 dilution; Cell Signaling Technology), KYN (H00008942-B01; 1:1,000 dilution; NOVUS Biologicals), TDO2 (H00006999-B01P; 1:1,000 dilution; Abnova), TOPO1 (C-21; 1:100 dilution; Santa Cruz Biotechnology, Inc.), and  $\alpha$ -tubulin (clone B-5-1-2; 1:30,000 dilution; Sigma Aldrich). Following secondary antibody incubation, results were detected using Western Lighting Chemiluminescence Reagent Plus (Perkin Elmer). Densitometry quantification was performed using Image Studio Lite



Version 3.1 and reported as a ratio compared with  $\alpha$ -tubulin as a loading control.

### Immunohistochemistry

Formalin-fixed, paraffin-embedded (FFPE) tissue sections (5  $\mu$ m) were heat immobilized onto glass slides and deparaffinized in a series of xylenes and graded ethanols. Antigens were heat retrieved in 10 mmol/L Tris, 1 mmol/L EDTA, pH 9.0 solution, and blocked for endogenous peroxidase followed by 10% normal goat serum prior to antibody incubation. Tris-buffered saline with 0.05% Tween (TBST) was used for all washes. TDO2 antibody (Abnova; # H00006999-B01P) was incubated overnight at room temperature at 1:200 in TBST, and AhR antibody (CST #13790) was incubated for 1 hour at room temperature at 1:50 in TBST. Envision polymer (Dako) was used for detection, followed by 3,3'-diaminobenzidine (Dako), and slides were counterstained with dilute hematoxylin.

### Gene expression array analysis

BT549 cells were grown in either attached conditions or forced-suspension conditions on poly-HEMA-coated plates in quadruplicate for 24 hours. RNA was harvested at 24 hours using Trizol method, and hybridized onto Affymetrix Human Gene 1.0ST arrays at the University of Colorado Denver Genomics and Microarray Core, following the manufacturer's instruction.

Microarray analysis was performed using Partek Genomics Suite (Partek, Inc.) and Ingenuity Pathway Analysis software (Qiagen, Inc.). One-way ANOVA analysis was performed to determine differentially expressed genes between the two treatment groups (attached vs. suspended). Fold-change cutoff was 1.5, and significance cutoff was  $P < 0.05$ . Significantly differentially expressed genes were imported to Ingenuity for pathway analysis, including identification of altered canonical pathways.

### shRNA experiments

High-titer shRNA lentiviral transduction particle suspensions were obtained from the Functional Genomics Facility at the University of Colorado (Boulder, CO). BT549 or MDA-MB-231 cells were plated at 50% confluence in 60-mm tissue culture-treated dishes in 4 mL of media. Twenty-four hours after plating, 100  $\mu$ L of the lentiviral suspension and 8  $\mu$ g/mL polybrene (Sigma-Aldrich) were added to the tissue culture media. After 24 hours, viral media were replaced with regular tissue culture media and cells were incubated for an additional 24 hours before the commencement of puromycin selection (1  $\mu$ g/mL). Knock-downs were confirmed by Western blot, and all experiments were done within three passages of puromycin selection. For shRNA sequences, see Supplementary Methods.

### Luciferase reporter activity

The AhR luciferase reporter, generously provided by Dr. Michael Denison (University of California-Davis), and NF- $\kappa$ B luciferase reporter, generously provided by Dr. Rebecca Schweppe (University of Colorado-Anschutz Medical Campus), were transiently transfected along with SV40 Renilla in TNBC cells using Lipofectamine (Life Technologies). Transfected cells were then plated in a 24-well plate at a density of  $5 \times 10^5$  cells per well in either a control or poly-HEMA-coated well and incubated at 37°C for 24 hours. Following incubation, reporter activation was determined using the Dual-Luciferase Reporter Assay System (Promega) according to the manufacturer's protocol. Briefly, cells

were lysed for 15 minutes at room temperature using  $1 \times$  passive lysis buffer. Lysed cells were collected and centrifuged at 14,000 rpm for 15 minutes at 4°C to eliminate cell debris. The supernatant was used immediately or diluted with  $1 \times$  passive lysis buffer for determination of luciferase activity. For analysis, AhR or NF- $\kappa$ B reporter activity was normalized to SV40 reporter activity to control for differences in transfection efficiency.

### Measurement of anoikis

Soft-agar assays were performed in 0.5% bottom and 0.25% top-layer agar (Difco Agar Noble; BD Biosciences). Media with treatment were refreshed every 4 days.

Caspase-3/7 activity was measured using a Caspase-Glo 3/7 Assay Kit (Promega) according to the manufacturer's protocol. Briefly,  $8 \times 10^4$  TNBC cells were plated in a clear bottom, white-walled 96-well plate coated with poly-HEMA. The cells were incubated for 48 hours at 37°C. Following incubation, each well was mixed with Caspase-Glo 3/7 reagent in equal volume to the culture medium. The 96-well plate was then covered in foil, shook for 30 seconds, and incubated at room temperature for 30 minutes. Luminescence was determined by luminometer, measuring luminescence after 1 second.

### In vivo metastasis experiment

Tail-vein injection experiments were approved by the University of Colorado Institutional Animal Care and Use Committee [IACUC protocol—83612(10)1E]. All animal experiments were conducted in accordance with the NIH Guidelines of Care and Use of Laboratory Animals. Prior to tail-vein injection, luciferase-expressing MDA-MB-231 cells were grown in forced suspension conditions for 48 hours in the presence of either vehicle control or 10  $\mu$ mol/L 680C91. A total of 250,000 viable cells, as determined by trypan blue staining, were then injected into the tail vein of NOD.CB17-Prkdc<sup>scid</sup>/J (NOD/SCID) mice (The Jackson Laboratory). Following injection, luminescence was monitored every 7 days for a total of 4 weeks. At the completion of this experiment, lungs of 5 mice from each group were imaged by IVIS *ex vivo*. Lungs from the remaining 5 mice per group were formalin-fixed and paraffin-embedded for IHC.

### Proliferation, migration, and invasion

Proliferation assays were performed using the Incucyte ZOOM live-cell imaging system (Essen BioSciences). Migration scratch wound assays were performed per the manufacturer's instructions and scanned with the Incucyte ZOOM apparatus (Essen BioSciences). Trans-well invasion assays were performed with BD BioCoat Matrigel Invasion Chambers (BD Biosciences) per the manufacturer's protocol.

### Statistical analysis

Statistical analysis was performed using GraphPad Prism 5. Student *t* test, ANOVA with Tukey post-hoc test, and two-way ANOVA with Bonferroni multiple comparison test were used as noted. *P* values are denoted as follows: \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.001$ ; \*\*\*\*,  $P < 0.0001$ ; ns, not significant.

## Results

### Multiple enzymes of the kynurenine pathway are upregulated in detached TNBC cell lines

To model loss of attachment *in vitro*, we grew breast cancer cells on tissue culture plates coated with poly(2-hydroxyethyl



methacrylate) (poly-HEMA). We previously demonstrated that TNBC cells grown in forced suspension upregulate proteins involved in anoikis resistance and motility (30, 31). To identify additional pathways responsible for resistance to anoikis in an unbiased, global fashion, expression profiling was performed on the TNBC cell line BT549 grown in standard attached conditions for 24 hours as compared with the same number of cells grown in forced suspension for 24 hours in quadruplicate. In total, we identified 367 genes that change more than 1.5-fold in a statistically significant manner ( $P < 0.05$ , log-rank test). Of these genes, 217 (59%) were upregulated and 150 (41%) were downregulated in suspended cells (Supplementary Fig. S1A). Ingenuity Pathway Analysis of these data revealed two related pathways strongly upregulated in suspension: tryptophan catabolism and AhR signaling (Supplementary Fig. S1B and S1C).

Figure 1A shows unsupervised hierarchical clustering of the top 35 genes significantly upregulated by at least 2-fold in suspended compared with attached conditions ( $P < 0.05$ , log-rank test). Among the genes most highly upregulated in suspension were two enzymes involved in the kynurenine pathway, which converts tryptophan to  $\text{NAD}^+$ . The rate-limiting enzyme tryptophan 2,3-dioxygenase (*TDO2*) was the most highly upregulated gene in suspension, and the downstream enzyme kynureninase (*KYNU*) was also among the top fold-changing genes (Fig. 1A). To confirm our gene array data, we performed qRT-PCR for *TDO2* and *KYNU* in multiple breast cancer cell lines, including both luminal ( $\text{ER}^+$ ) and TNBC ( $\text{ER}^-$ ) lines, after 24 hours in suspension (Fig. 1B and C). In all three TNBC lines tested, *TDO2* and *KYNU* were significantly increased in suspension compared with attached culture. In the two  $\text{ER}^+$  breast cancer cell lines tested, expression of these genes trended slightly higher in suspension, but this change was not significant.

Western blot analysis of whole-cell extracts also demonstrated an increase in *TDO2* and *KYNU* proteins in TNBC cell lines (MDA-231, BT549, and SUM159) grown in suspension for 24 hours (Fig. 1D; and Supplementary Fig. S3A). The increase in *TDO2* protein was confirmed by IHC in BT549 cells grown in suspension for 48 hours compared with cells grown in the attached condition (Fig. 1E).

Global metabolomic profiling of intracellular and secreted metabolites from BT549 cells grown in standard attached conditions or in forced suspension for 24 hours was also performed. Two intermediate products of the kynurenine pathway, Kyn and formylkynurenine, were the intracellular metabolites with the highest fold-change increase in suspension. Among secreted metabolites, kynurenine had the third-highest fold-change increase (Supplementary Fig. S2). Together with the gene expression data, this demonstrates that the kynurenine pathway is strongly upregulated in TNBC cells upon loss of attachment.

Using HPLC to verify the metabolomic profiling data, we found that secreted Kyn levels were more than 2-fold higher in conditioned media from BT549 cells in forced suspension for 48 hours than in media from the same number of cells in the attached condition (Fig. 1F). Furthermore, addition of the *TDO2*-specific inhibitor 680C91 to cells in suspension completely prevented the increase in secreted Kyn, demonstrating that increased secretion of Kyn in TNBC cells in suspension is dependent on *TDO2* activity (Fig. 1F). *TDO2* mRNA was expressed at a higher copy number at baseline than *IDO1* in two of the three TNBC cell lines tested, and *TDO2* expression increased more strongly in suspension than *IDO1* (Supplementary Fig. S3B).

### Aryl hydrocarbon receptor expression and activity are increased in suspension

In addition to the components of the kynurenine pathway, *AhR* and *AhR*-regulated genes were also identified among the genes most highly upregulated by TNBC cells in suspension (Fig. 1A and Supplementary Fig. S4). Increased *AhR* mRNA expression in suspended compared with attached TNBC cells was confirmed by qRT-PCR. Similar to *TDO2* and *KYNU*, *AhR* was not significantly increased in  $\text{ER}^+$  cell lines in suspension (Fig. 2A). Furthermore, *AhR* protein was increased in all three TNBC cell lines tested following 24 hours in suspension as tested by Western blot (Fig. 1D; Supplementary Fig. S3A), and in BT549 cells grown in suspension for 48 hours as measured by IHC (Fig. 2C). In attached SUM159 cells, while *AhR* protein was found in both the cytoplasmic and nuclear fractions, after 48 hours in forced suspension, *AhR* protein was almost exclusively nuclear (Fig. 2B). Because *AhR* nuclear translocation and transcriptional activity are known to be ligand-mediated (32), the movement of *AhR* to the nucleus in suspension suggests that the receptor is ligand-bound and transcriptionally active.

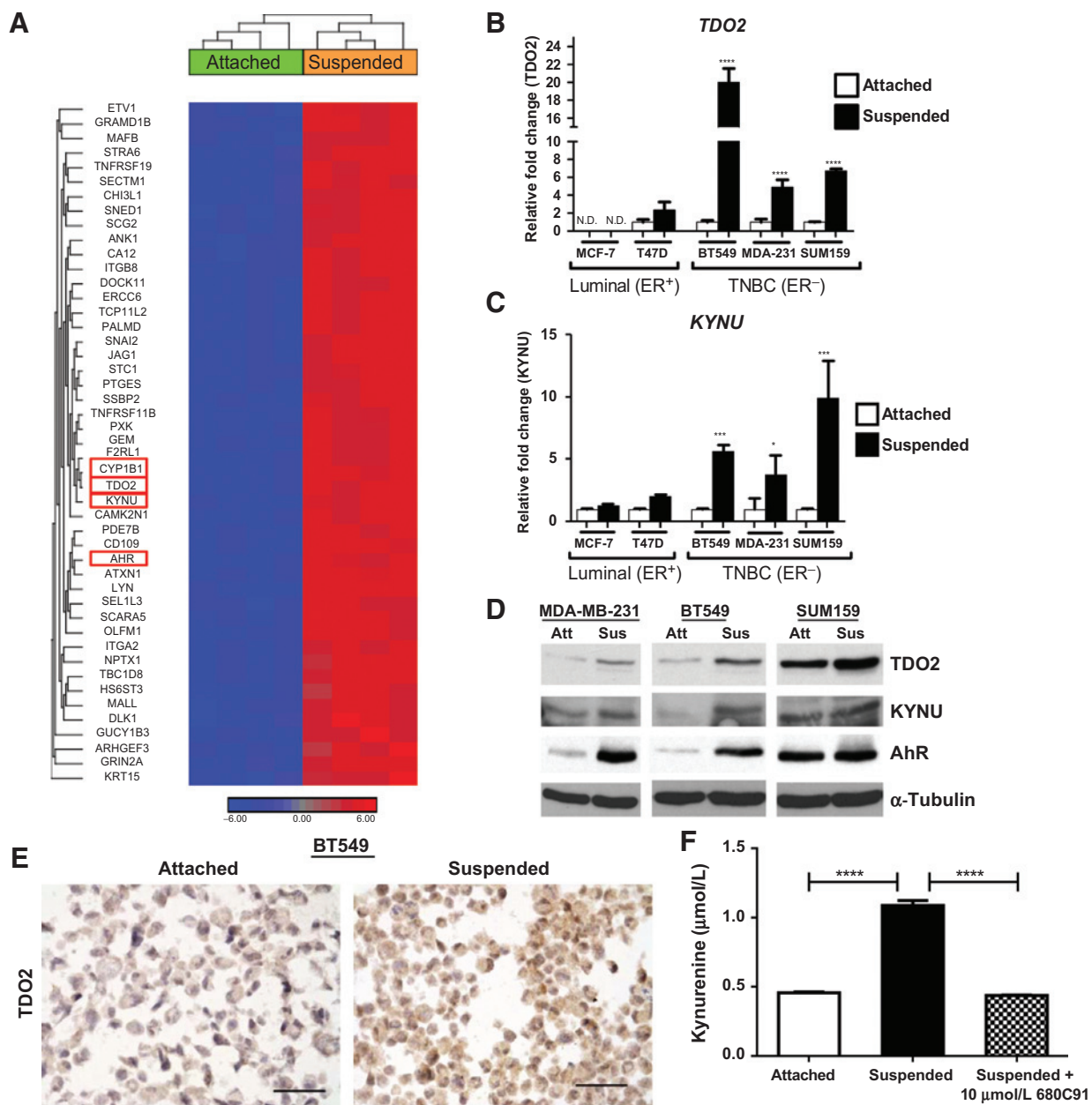
We assessed *AhR* transcriptional activity using a luciferase reporter containing six dioxin response elements (DRE), the consensus *AhR*-binding site. In BT549 and MDA-231 cells, luciferase activity was upregulated more than 2-fold in suspended compared with attached cells (Fig. 2D). The *AhR* antagonist  $\alpha$ -naphthoflavone inhibited the suspension-mediated increase in luciferase activity, demonstrating specificity of the reporter construct for *AhR* activity (Fig. 2D; Supplementary Fig. S3C).

### *TDO2*-mediated kynurenine production activates *AhR*

Kyn is an endogenous ligand for *AhR* (12, 33–35), suggesting a potential relationship between increased kynurenine pathway enzyme expression and increased *AhR* activity in TNBC cells in suspension. First, we tested whether inhibition of *TDO2* and consequent Kyn production could prevent *AhR* activation in suspended TNBC cells. Treatment with 10  $\mu\text{mol/L}$  680C91, which prevented the suspension-induced increase in Kyn, significantly decreased *AhR* activity in BT549 and MDA-MB-231 cells in suspension (Fig. 2E; Supplementary Fig. S3D). Furthermore, the suspension-mediated increase in *AhR* target genes *CYP1A1* and *CYP1B1* was abrogated by both the *AhR* inhibitor CH-223191 and the *TDO2* inhibitor 680C91 (Fig. 2F), linking *TDO2* activity to *AhR* transcriptional activity. Addition of 100  $\mu\text{mol/L}$  exogenous Kyn also resulted in a significant increase in *AhR* luciferase reporter activity in TNBC cells in both attached and suspended culture conditions (Fig. 2G), demonstrating that Kyn is able to activate *AhR* in TNBC cells.

### Inhibition of *TDO2* and *AhR* decreases TNBC anoikis resistance, proliferation, migration, and invasion

Because *TDO2* and *AhR* were upregulated by TNBC cells in suspension, we next tested their functional importance by assessing whether pharmacologic inhibition or genetic knockdown could reduce anchorage-independent growth in soft agar or increase sensitivity of TNBC cells to anoikis in forced suspension culture. BT549 and SUM159PT cells were pretreated in the attached condition with vehicle, the *TDO2* inhibitor 680C91, or the *AhR* antagonist CH-223191 for 24 hours. Then an equal number of cells were plated in soft agar in the continued presence of treatment. Both 680C91 and CH-223191 significantly decreased anchorage-independent growth of BT549 and SUM159 cells in soft

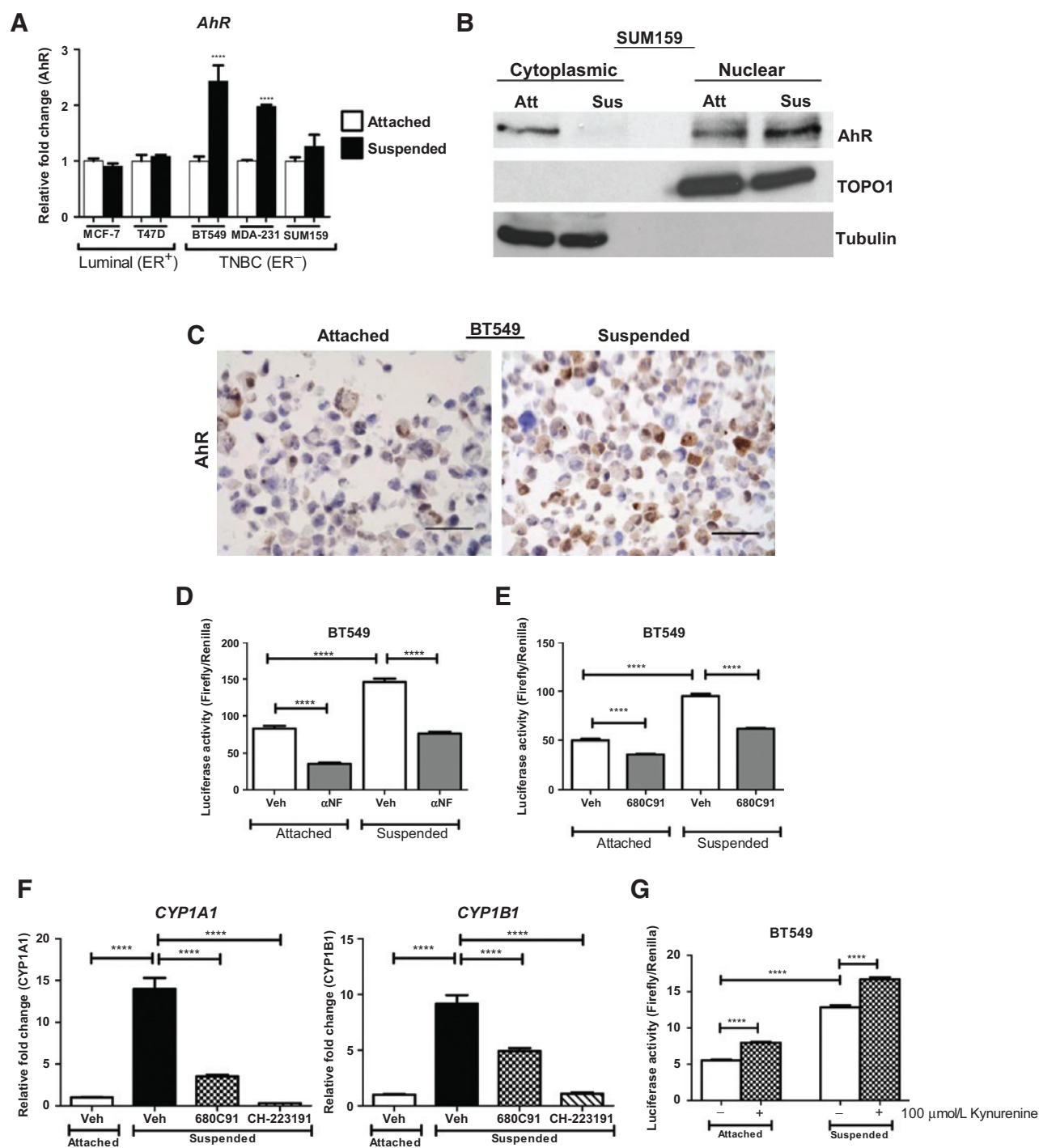
**Figure 1.**

Kynurenine pathway components and activity are increased in TNBC cells in suspension and suppressed by TDO2 inhibition. A, BT549 cells were grown in either attached or forced-suspension conditions on poly-HEMA-coated plates in biologic quadruplicate for 24 hours. RNA was harvested at 24 hours and hybridized onto Affymetrix Human Gene 1.0ST array. Gene expression profile analysis identified genes significantly upregulated in suspension culture ( $P < 0.05$ ). Those involved in tryptophan metabolism and AhR signaling are highlighted in red. B and C, relative TDO2 (B) and KYNU (C) mRNA levels were determined by qRT-PCR in luminal and TNBC cell lines grown in attached or suspended culture for 24 hours. D, Western blot for TDO2, KYNU, and AhR protein in three TNBC cell lines following 24 hours of attached or suspended culture. E, TDO2 protein detected by IHC in BT549 cell pellets grown in attached or suspended culture for 48 hours. Bar, 50 μm. F, secreted kynurenine levels, as measured by HPLC, from BT549 cells grown for 48 hours in the attached versus suspended conditions or suspended plus 10 μmol/L 680C91 (a specific TDO2 inhibitor). \*,  $P < 0.05$ ; \*\*\*,  $P < 0.001$ ; \*\*\*\*,  $P < 0.0001$  by ANOVA with Bonferroni multiple comparison test.

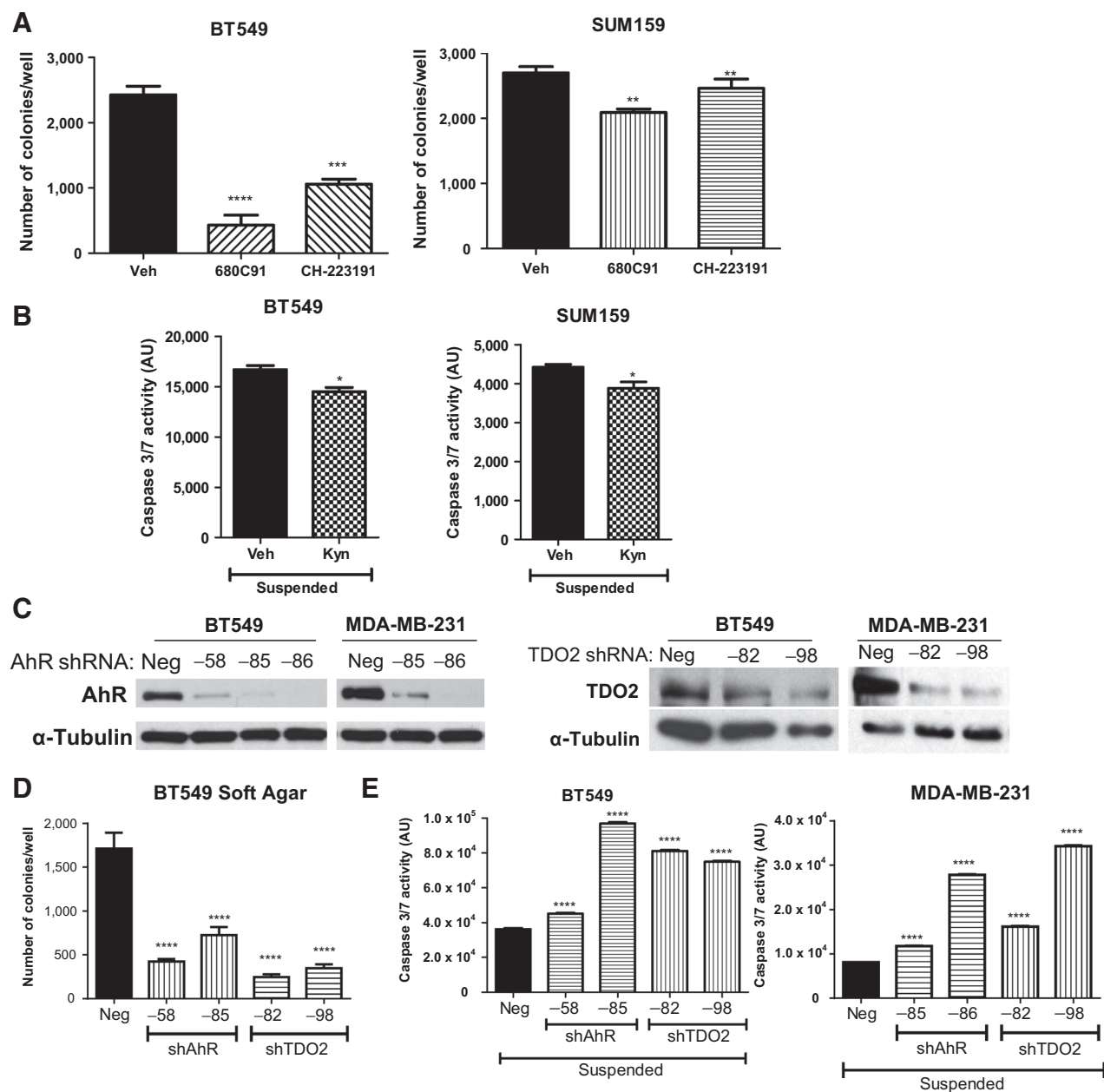
agar (Fig. 3A). To test the effect of Kyn on anoikis resistance, we treated cells in forced suspension culture for 48 hours with Kyn and found that this significantly decreased apoptosis as measured by cleaved caspase activity compared with vehicle control treatment (Fig. 3B). We then performed knockdown of TDO2 and AhR using two shRNA constructs each, and decreased protein expression was

confirmed by Western blot (Fig. 3C). Knockdown of either TDO2 or AhR also significantly decreased growth of BT549 cells in soft agar (Fig. 3D). Knockdown of either TDO2 or AhR also significantly increased apoptosis in BT549 and MDA-MB-231 cells grown in suspension for 48 hours (Fig. 3E), demonstrating that TDO2 and AhR promote survival in anchorage-independent conditions.

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**Figure 2.**

AhR expression and localization are affected by suspension culture, and kynurenine activates AhR in TNBC cells in suspension. Breast cancer cell lines were plated on uncoated (attached) or poly-HEMA-coated (suspended) tissue culture plates for 24 hours. A, relative *AhR* levels measured by qRT-PCR in ER<sup>+</sup> or TNBC cells in attached or suspended culture for 24 hours. B, Western blot for AhR levels in nuclear and cytoplasmic fractions of SUM-159 cells in attached or suspended culture for 24 hours. C, AhR protein as detected by IHC in BT549 cell pellets grown in attached or suspended culture for 48 hours. Bar, 50  $\mu$ m. D and E, AhR reporter activity in BT549 cells in attached or suspended culture for 24 hours treated with vehicle [0.1% MeOH:EA (3:1); D] or 10 mmol/L  $\alpha$ -Naphthoflavone (AhR antagonist) or vehicle (0.1% DMSO; E) or 10  $\mu$ mol/L 680C91 (TDOi). F, relative expression of AhR target genes by qRT-PCR in BT549 cells in attached or suspended culture treated with vehicle, 680C91, or CH-223191 (AhR antagonist) for 24 hours. G, AhR reporter activity in BT549 cells in attached or suspended culture for 24 hours treated with vehicle ( $2.5 \times 10^{-4}$  M HCl) or crystalline kynurenine (100  $\mu$ mol/L). \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; \*\*\*\*,  $P < 0.0001$  by ANOVA with Bonferroni multiple comparison test.

**Figure 3.**

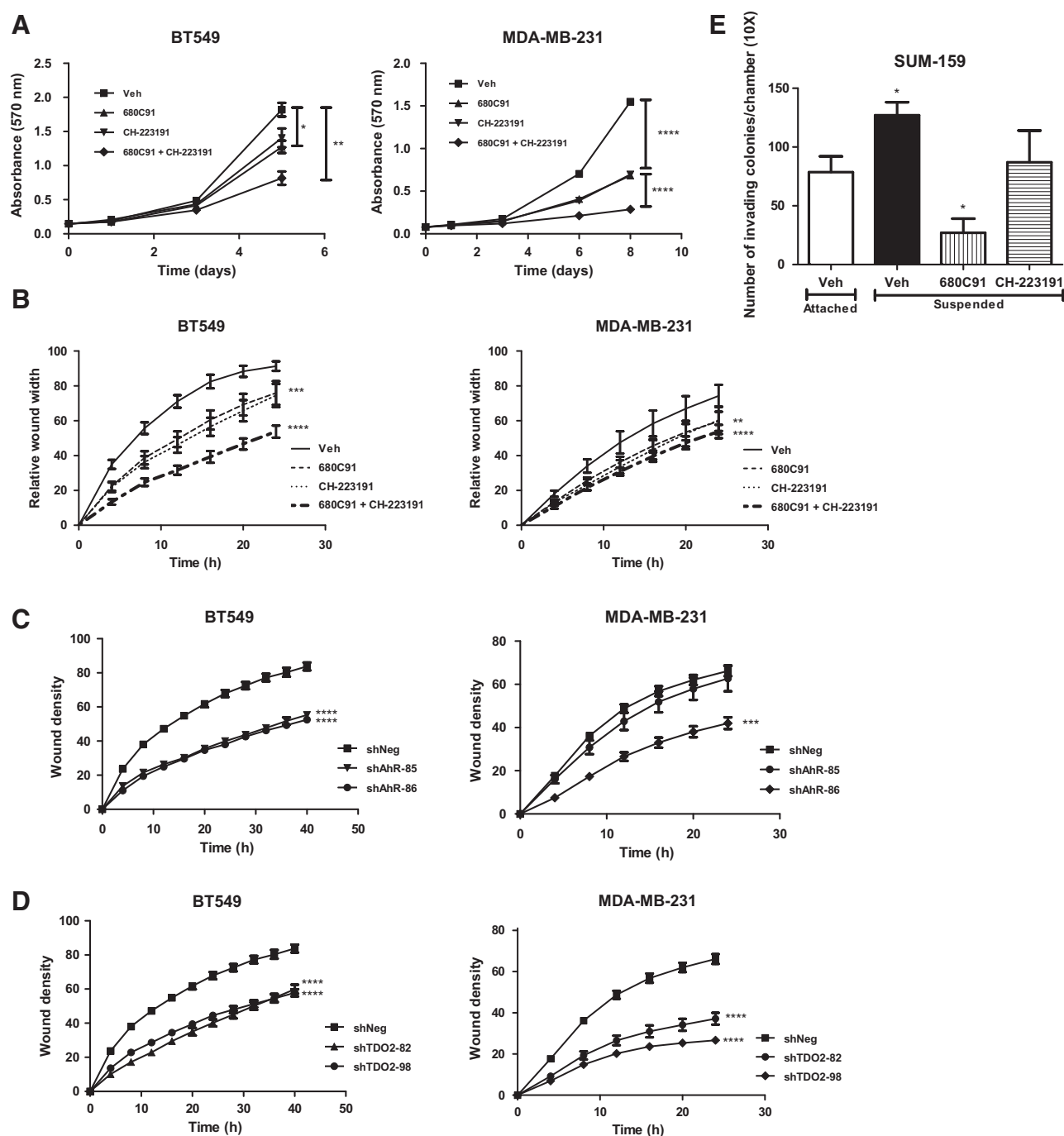
Targeting the kynurenine pathway or AhR increases cell death of TNBC cells in suspension. A, anchorage-independent growth of SUM159 or BT549 cells pretreated with vehicle (0.01% DMSO), 10  $\mu$ M/L 680C91 (TDOi), or 10  $\mu$ M/L CH-223191 (AhR antagonist) for 24 hours, then plated in 0.25% soft agar where respective treatments were maintained for 18 to 21 days. B, caspase 3/7 activity in TNBC cells (BT549 and SUM159) treated with either vehicle control or 100  $\mu$ M/L kynurenine while cultured in suspension for 48 hours. C, Western blot of TDO2 or AhR in attached BT549 or MDA-MB-231 cells following transduction with nontargeting shRNA (shNEG) or shRNA constructs targeting AhR or TDO2. D, anchorage-independent soft-agar growth of BT549 cells with knockdown of TDO2 or AhR grown in soft agar for 14 days. E, caspase 3/7 activity in BT549 or MDA-MB-231 cells transduced with nontargeting shRNA (shNEG) or shRNA constructs targeting AhR or TDO2, and plated in poly-HEMA-coated plates for 48 hours. \*\*,  $P < 0.01$ ; \*\*\*\*,  $P < 0.0001$  by ANOVA with Bonferroni multiple comparison test.

To determine if TDO2 and AhR also mediate growth of TNBC cells, MDA-MB-231 and BT549 cells were treated with 680C91 or CH-223191, and cell number was measured over time. Both inhibitors significantly decreased the number of cells when grown in traditional attached conditions, and the combination was more effective than either drug alone (Fig. 4A). MDA-MB-231, BT549, and SUM159 cells were also treated

with a range of 680C91 concentrations, and GI50 values were calculated as 20  $\mu$ M/L, 61  $\mu$ M/L, and 102  $\mu$ M/L, respectively (Supplementary Fig. S5). Knockdown of TDO2 or AhR similarly resulted in decreased growth of MDA-231 cells (Supplementary Fig. S6), demonstrating a role for TDO2 and AhR signaling in baseline growth of TNBC cells *in vitro* even in attached culture.



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**Figure 4.**

TDO2 and AhR inhibition decreases cell number, migration, and invasion of TNBC cells. BT549 and MDA-MB-231 cells (TNBC) were treated with either vehicle control (0.01% DMSO), 10  $\mu\text{mol/L}$  680C91 (TDOi), 10  $\mu\text{mol/L}$  CH-223191 (AhR antagonist), or in combination. A, crystal violet assay of BT549 and MDA-MB-231 cells treated for 8 days. B, scratch wound assay of BT549 and MDA-MB-231 cells treated in media containing 0.5% FBS. C and D, scratch wound assay of BT549 and MDA-MB-231 cells transduced with a nontargeting control (shNeg) or shRNAs targeting AhR (shAhR-85, shAhR-86; C) or TDO2 (shTDO2-82, shTDO2-98; D). E, Boyden chamber invasion assay of SUM-159 cells pretreated with either vehicle control (0.01% DMSO), 10  $\mu\text{mol/L}$  680C91 (TDOi), or 10  $\mu\text{mol/L}$  CH-223191 (AhR antagonist) in the attached or suspended conditions for 24 hours, then seeded into the upper chamber and allowed to invade through a Matrigel-coated membrane. \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; \*\*\*\*,  $P < 0.0001$  by ANOVA with Bonferroni multiple comparison test.

In addition, pharmacologic inhibition of TDO2 and AhR each significantly reduced migration of MDA-MB-231 and BT549 cells in a scratch wound assay, and again the combination was more effective than either inhibitor alone (Fig.

4B). Knockdown of either TDO2 or AhR recapitulated this effect, significantly diminishing migration in the scratch wound assay in both BT549 and MDA-231 cells (Fig. 4C and D).

To test whether the increased TDO2 and AhR expression observed in cells grown in suspension affects invasive capacity, SUM159PT cells were grown for 48 hours either in the attached condition, or in forced suspension culture with or without addition of 10  $\mu\text{mol/L}$  of the TDO2 inhibitor 680C91 or the AhR inhibitor CH-223191. After 24 hours, 25,000 viable cells were plated in a Matrigel-coated transwell invasion chamber with continuous treatment. Cells grown in suspension for 24 hours were significantly more invasive than control cells grown in the attached condition (Fig. 4E). The addition of the TDO2 inhibitor 680C91 during suspension culture greatly decreased the invasive capacity of viable cells. Although the cells treated with the AhR inhibitor showed decreased invasion, this effect was not statistically significant (Fig. 4E).

#### NF- $\kappa$ B regulates expression of kynurenine pathway genes and AhR in TNBC cells

We previously demonstrated that NF- $\kappa$ B activity is substantially increased in TNBC cells in suspension, but not in ER<sup>+</sup> cells (31). Here, we again demonstrate increased NF- $\kappa$ B activity in TNBC cells in suspension as measured by an NF- $\kappa$ B luciferase reporter, and this increase is not observed using a reporter containing a mutated binding site (Fig. 5A). Interestingly, a recent global profiling study of ovarian cancer *in vivo* found that knockdown of I $\kappa$ B-epsilon, a positive regulator of NF- $\kappa$ B activity, decreased expression of TDO2 and KYNU, suggesting NF- $\kappa$ B regulation of these genes (36); however, the regulatory mechanism was not directly tested.

To investigate the link between increased NF- $\kappa$ B activity, TDO2, and KYNU, we treated BT549 cells with a cocktail of TNF $\alpha$  and IL1 $\beta$  to activate NF- $\kappa$ B and found that expression of TDO2, KYNU, and AhR (Fig. 5B), as well as the NF- $\kappa$ B target genes IL6 and IL8 (data not shown), was significantly increased at 24 hours. Next, we tested whether inhibition of NF- $\kappa$ B in suspension could reduce upregulation of Kyn pathway genes and AhR in TNBC cells grown in suspension. Upstream inhibition of NF- $\kappa$ B with the I $\kappa$ B inhibitor PS1145 significantly reduced the suspension-induced increase in TDO2, KYNU, and AhR (Fig. 5C), as well as the NF- $\kappa$ B target gene IL6 (not shown), demonstrating that NF- $\kappa$ B activity mediates the increased TDO2, KYNU, and AhR expression in TNBC cells in suspension. Lastly, a constitutively active form of I $\kappa$ B, a negative regulator of NF- $\kappa$ B, was expressed in BT549 cells, and this ablated the suspension-induced increase in TDO2, KYNU, and AhR (Fig. 5D). These data demonstrate that the increased transcription of TDO2, KYNU, and AhR in suspension is specifically induced by and dependent on NF- $\kappa$ B in TNBC.

#### TDO2 inhibition decreases metastatic colonization *in vivo*

To test the potential contribution of TDO2 activity to the metastatic capacity of TNBC cells *in vivo*, we grew luciferase-expressing MDA-MB-231 cells in forced suspension conditions for 48 hours in the presence of either vehicle control or the TDO2 inhibitor 680C91. A total of 250,000 viable cells, as determined by trypan blue staining, were then injected into the tail vein of NOD/SCID mice, and luminescence was monitored over time. Seven days after injection, mice that received vehicle-treated cells had significantly higher luminescence, and this statistically significant difference was maintained throughout the experiment (Fig. 6A and B). At the conclusion of the experiment at day 28 after injection, lung luminescence was

measured *ex-vivo*, and lungs from mice that received vehicle-treated cells had significantly higher luminescence compared with mice that received cells treated with the TDO2 inhibitor (Fig. 6C). A significant decrease in the number of metastatic nodules in the lungs from mice receiving cells treated with 680C91 was also observed by hematoxylin and eosin (H&E; Fig. 6D). Together, these data demonstrate that TDO2 inhibition decreases the ability of TNBC cells to successfully metastasize following tail vein injection *in vivo*.

#### TDO2 is more highly expressed in ER<sup>-</sup> than ER<sup>+</sup> breast cancer and correlates with poor prognosis

To determine the clinical relevance of our findings, we examined TDO2 expression in primary TNBC samples by IHC. As shown in Fig. 7A, primary TNBC were strongly positive for TDO2 expression. We evaluated TDO2 gene expression in patient samples using publicly available gene expression microarray datasets from Oncomine. The Curtis and colleagues dataset (37) had the largest patient population ( $n = 1,998$ ) and was therefore selected for analysis. Notably, TDO2 was the sixth most highly overexpressed gene in breast carcinoma compared with normal breast tissue (top 1% of overexpressed genes,  $P < 0.0001$ ; Fig. 7B). TDO2 gene expression was significantly higher in ER-negative breast tumors compared with ER-positive tumors ( $P < 0.0001$ ; Fig. 7C). High TDO2 expression was also associated with increasing grade ( $P < 0.001$ ; Fig. 7D), supporting the hypothesis that tumors with high TDO2 expression may have increased metastatic potential. Similarly, IDO1 was overexpressed in breast carcinoma compared with normal breast tissue, although it was not as highly overexpressed as TDO2 (Supplementary Fig. S7A). IDO1 was also more highly expressed in ER<sup>-</sup> than ER<sup>+</sup> breast tumors in both the Curtis and TCGA datasets (Supplementary Fig. S7B and S7C).

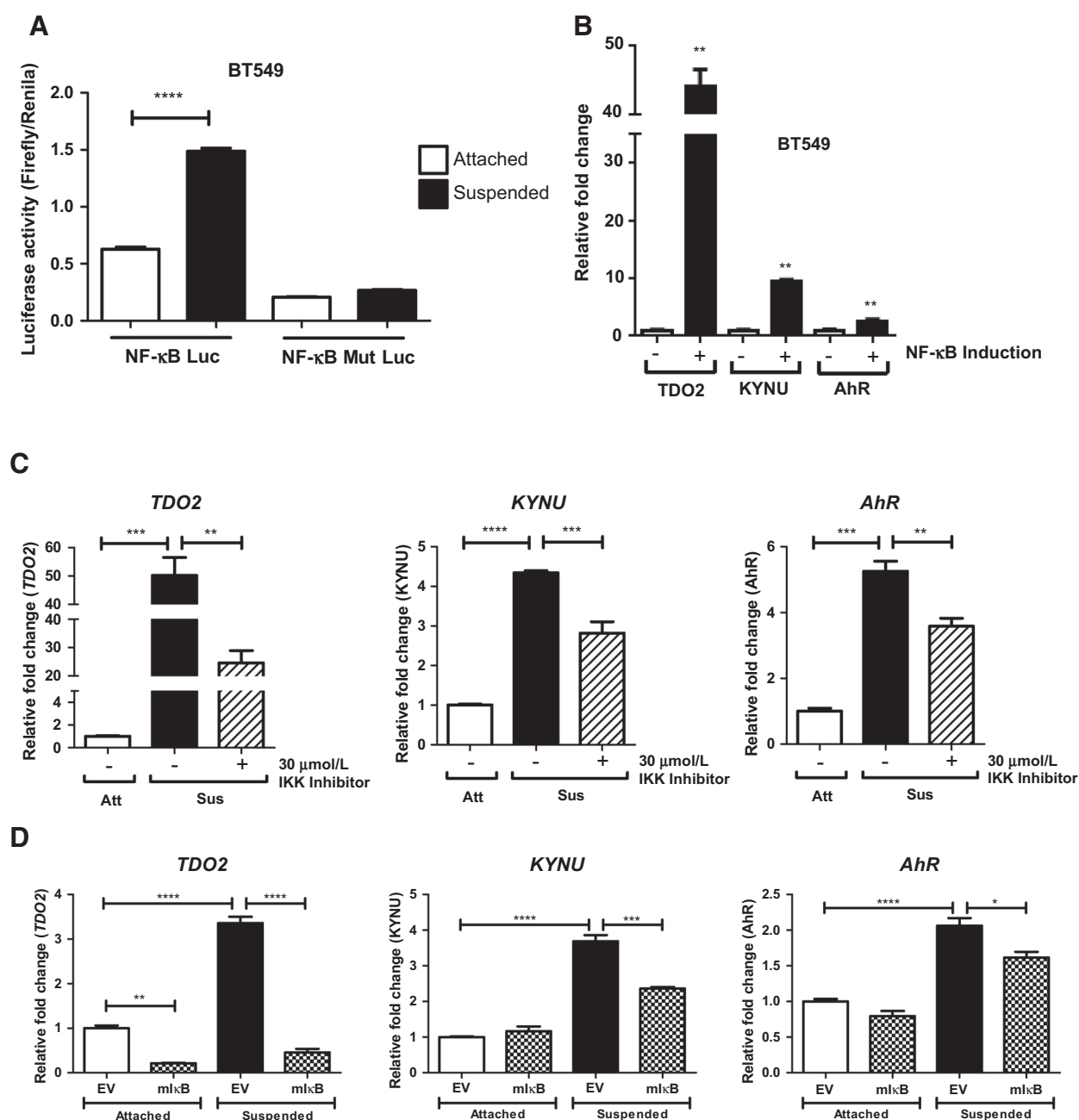
Finally, patients whose tumors had above-median TDO2 expression had approximately 3 years shorter overall survival when compared with those with below-median TDO2 expression (median overall survival 10.62 years vs. 13.31 years, respectively;  $P = 0.0002$ , log-rank test; Fig. 7E), suggesting that TDO2 expression may contribute to tumor progression and poor prognosis. Similarly, patients with above-median IDO1 expression had a shorter survival than those with lower IDO1, but the difference in survival was not as large (Supplementary Fig. S7D). This further supports the importance of targeting TDO2 in breast cancer patients, possibly in combination with IDO1, rather than targeting IDO1 alone as in current clinical trials.

## Discussion

TNBC has no effective targeted therapies and the worst prognosis of the breast cancer subtypes, due in part to its propensity for rapid metastasis (3). Indeed, patients with metastatic TNBC have a median survival of only 13 months (5). However, few studies have examined pathways that support anoikis resistance in TNBC. In this study, global gene expression profiling revealed the novel finding that TNBC cells in forced suspension upregulate two related pathways: AhR signaling and the kynurenine pathway of tryptophan catabolism. Indeed, global metabolomic profiling identified formylkynurenine and Kyn as the most highly upregulated intracellular metabolites, and Kyn as the third most-highly



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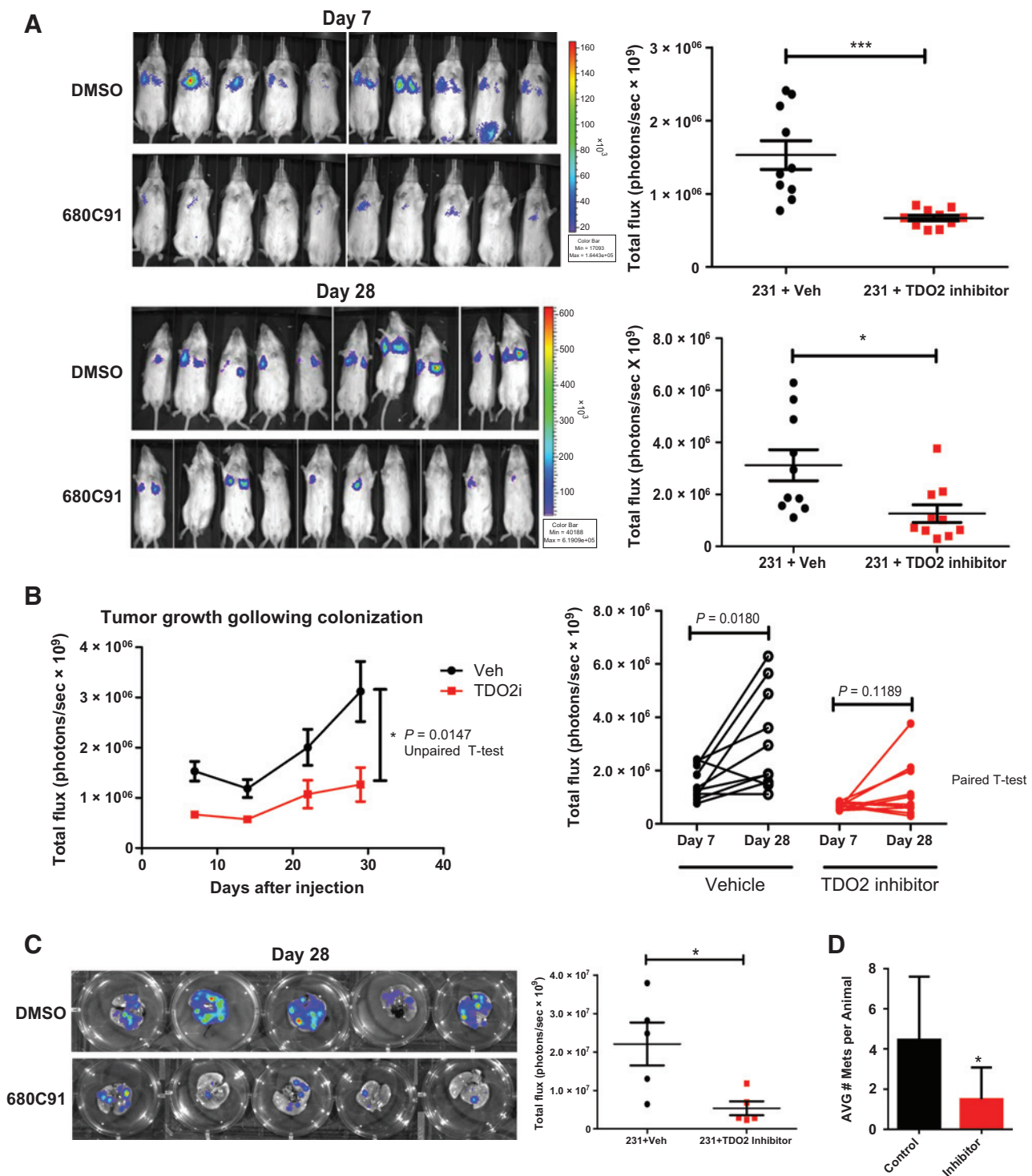
**Figure 5.**

NF-κB regulates expression of TDO2, KYNU, and AhR in TNBC cells in suspension. A, activity of an NF-κB (left) or mutated NF-κB (right) luciferase reporter in BT549 cells in attached or suspended condition for 24 hours. B, relative expression of *TDO2*, *KYNU*, and *AhR* by qRT-PCR in BT549 cells treated with vehicle or TNFα (10 ng/mL) plus IL1β (10 ng/mL) for 24 hours. C, relative expression of *TDO2*, *KYNU*, and *AhR* by qRT-PCR in BT549 cells grown in attached or suspended culture treated with vehicle (0.01% DMSO) or 30 μmol/L PS1145 (IKK inhibitor) for 24 hours. D, relative expression of *TDO2*, *KYNU*, and *AhR* as measured by qRT-PCR in BT549 cells grown in attached or suspended culture and expressing a stable empty-vector (EV) or mutant (constitutively active) IκB expression vector. \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.001$ ; \*\*\*\*,  $P < 0.0001$  by ANOVA with Bonferroni multiple comparison test.

increased secreted metabolite in conditioned media from cells in forced suspension.

Based on our data demonstrating that TNBC cells in suspension upregulate AhR and TDO2, increase secretion of Kyn, and rely on TDO2 and AhR for anchorage-independent growth and invasive potential, we hypothesized that suspended TNBC cells

utilize an autocrine signaling loop in which Kyn activates AhR to support anoikis resistance and anchorage-independent growth, as well as migration and invasion (Fig. 7F). Central to this model is upregulation of TDO2, a key enzyme in the Kyn pathway. Our observation that pharmacologic inhibition of TDO2 decreased the ability of MDA-MB-231 cells to form lung

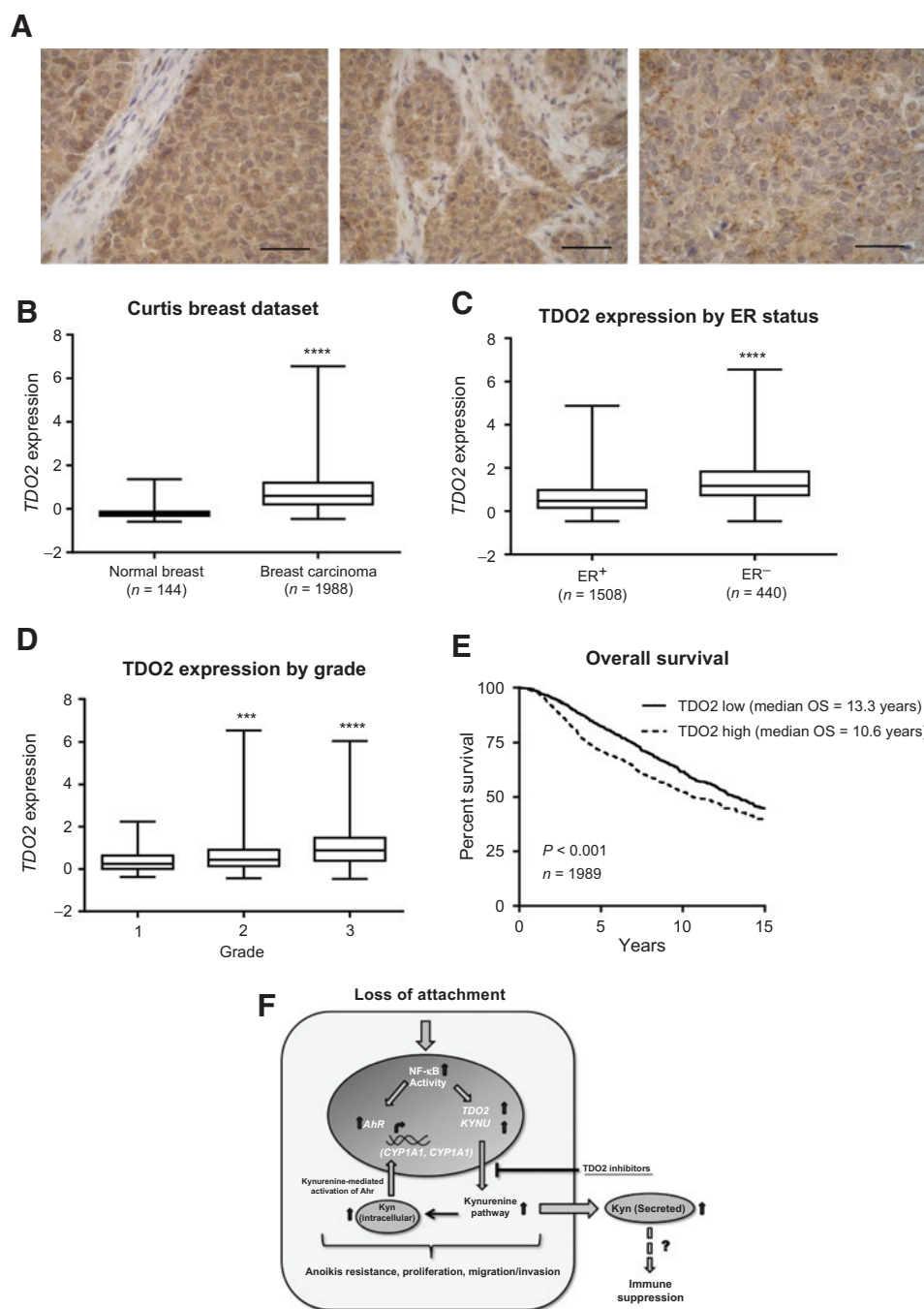
**Figure 6.**

Targeting TDO2 decreases colonization and outgrowth of TNBC cells *in vivo*. Luciferase-expressing MDA-MB-231 cells were pretreated for 48 hours in forced-suspension culture with either vehicle (0.01% DMSO) or 10  $\mu\text{mol/L}$  680C91 prior to tail vein injection. A and B, metastatic colonization was measured by whole animal IVIS imaging at 7 days after injection and weekly thereafter. C, *ex vivo* imaging of lungs 28 days after injection. D, quantification of lung metastases based on H&E of FFPE lung sections. \*,  $P < 0.05$ ; \*\*\*,  $P < 0.001$  by two-tailed *t* test.

metastases *in vivo* suggests that TDO2 inhibition represents a novel opportunity for targeted therapy to inhibit TNBC metastasis.

Further, we show that the Kyn pathway components TDO2 and KYNU, as well as AhR, are regulated by the transcription factor NF- $\kappa\text{B}$  in suspension. We previously identified a different

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**Figure 7.**

TDO2 expression in breast cancer clinical samples. A, TDO2 staining in primary TNBC patient samples; bar, 50  $\mu$ m. B–E, using the Curtis breast cohort, TDO2 expression was compared between normal breast and invasive ductal carcinoma (B). TDO2 gene expression in breast tumor samples divided by ER<sup>+</sup> versus ER<sup>−</sup> (C) or by tumor grade (D). E, patients were then split into TDO2-high or -low groups based on median TDO2 gene expression, and overall survival was plotted. F, a model of TDO2-AhR signaling in TNBC cells. NF- $\kappa$ B activity increases in suspension, increasing expression of TDO2, which leads to increased Kyn (both intracellular and secreted), which subsequently activates AhR and promotes anois resistance, migration, and invasion.  $P < 0.001$  by log-rank test.

NF- $\kappa$ B-regulated autocrine signaling loop involving the neurotrophic tyrosine kinase, type 2 (NTRK2 or TrkB), and the TrkB ligand neurotrophin 3 (NTF3) that promotes survival of TNBC cells in suspension (31), suggesting that increased NF- $\kappa$ B signaling in suspended cells supports multiple prometastatic attributes of TNBC.

In addition to its autocrine role affecting growth, migration, and invasion of tumor cells that we describe here, the Kyn pathway may also facilitate metastasis through paracrine

signaling mechanisms involving suppression of immune surveillance. Specifically, TDO2-mediated Kyn production prevented T-cell-mediated immune rejection of the immunogenic P815 mouse tumor cell line in immunized mice (38). In dendritic cells, IDO1 activity can lead to the AhR-dependent generation of regulatory T cells via a coordinated increase in Kyn and decrease in tryptophan (33). Inhibition of IDO1 also sensitizes cancer cells to chemotherapy in a T-cell-dependent manner (39).

Multiple clinical trials testing the efficacy of Indoximod (1-methyl-D-tryptophan), an IDO1/2 inhibitor, are currently under way, including one in metastatic breast cancer in combination with conventional chemotherapies or immunotherapy (NCT01792050). These trials are based on the ability of IDO1/2 inhibition to reverse immune suppression and enhance the antitumor immune response. However, 1-MT does not target TDO2 (40). Importantly, our data combined with data mined from publicly available breast cancer clinical cohorts suggest that TDO2 may be the more relevant target in TNBC. Interestingly, the membrane protein responsible for tryptophan import into cells, LAT1, is also more highly expressed in TNBC than in other breast cancer subtypes (41). Although a recently published analysis of serum metabolite concentrations in breast cancer patients found no difference in tryptophan levels between ER<sup>+</sup> and ER<sup>-</sup> breast cancer patients, median levels of Kyn were significantly higher in patients with ER<sup>-</sup> tumors (42). Although this finding was attributed to IDO1 activity, we find that TDO2 is higher in ER<sup>-</sup> tumors than ER<sup>+</sup> in two publicly available datasets, and that TDO2 expression is higher than IDO1 at baseline in two of three TNBC cell lines tested and TDO2 increases more strongly in suspension in all cell lines. Lastly, and perhaps the strongest indicator of the relative role of TDO2 versus IDO1, blocking TDO2 activity completely prevented the detachment-induced increase in Kyn secretion observed in TNBC cells in suspension (Fig. 1F).

Collectively, our results suggest that in TNBC, TDO2 is an important contributor to Kyn production, and thus should be considered as a therapeutic target. High TDO2 in primary breast tumors from publicly available datasets correlates with shorter overall survival. TDO2 was significantly upregulated in metastases of leiomyosarcoma patients compared with primary tumors (43), and increased KYN expression is associated with metastasis of breast cancer cells to the lung and brain (44, 45). In addition to abrogating the autocrine protumorigenic effects of Kyn, TDO2 inhibition may also inhibit Kyn activation of AhR in immune cells and thereby reduce TNBC immune evasion, but this remains to be tested.

Although we demonstrate a significant increase in Kyn levels in TNBC cells in suspension, it is possible that the effects of TDO2 could be mediated in part through production of other intermediate molecules or NAD<sup>+</sup> production. However, AhR nuclear localization and transcriptional activity are clearly increased in TNBC in suspension culture, and other recent studies find that AhR affects growth and migration of breast cancer cells (27, 28) and castrate-resistant prostate cancer cells (46). Studies examining AhR expression in breast cancer primary tumors found that higher AhR expression correlates with good prognosis (47). However, our data suggest that AhR expression, nuclear localization, and transcriptional activity may be low until cells begin the metastatic process by detaching from the primary tumor. Thus, AhR in the primary tumor may not reflect its role in supporting the metastatic cascade.

The independent tumor-promoting roles of the Kyn pathway (particularly via IDO1) and AhR signaling have been recog-

nized separately for many years. Recently, the connection of these two pathways via Kyn serving as an AhR ligand was discovered (12, 33–35). Our study demonstrates that the TDO2-AhR signaling axis may be of particular importance in TNBC because it is strongly activated in this highly metastatic breast cancer subtype in response to detachment, and supports anchorage-independent survival and invasive capacity. The combination of protumorigenic autocrine effects and potential immunosuppressive paracrine signaling effects makes this pathway an attractive target for therapeutic intervention. Our findings that inhibition of the Kyn or AhR pathways decreased *in vitro* characteristics associated with multiple steps in the metastatic cascade, and that pharmacologic inhibition of TDO2 decreased TNBC metastasis *in vivo*, suggest that inhibitors of TDO2 may represent an exciting opportunity for targeted therapy for TNBC to decrease mortality from this aggressive breast cancer subtype.

### Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

### Authors' Contributions

Conception and design: N.C. D'Amato, T.J. Rogers, L.I. Greene, D.R. Cochrane, J.K. Richer

Development of methodology: T.J. Rogers, L.I. Greene, T.G. Nemkov, J.K. Richer

Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): N.C. D'Amato, L.I. Greene, D.R. Cochrane, N.S. Spoelstra, T.G. Nemkov, A. D'Alessandro, J.K. Richer

Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): N.C. D'Amato, T.J. Rogers, M.A. Gordon, L.I. Greene, N.S. Spoelstra, T.G. Nemkov, K.C. Hansen, J.K. Richer

Writing, review, and/or revision of the manuscript: N.C. D'Amato, T.J. Rogers, M.A. Gordon, N.S. Spoelstra, K.C. Hansen, J.K. Richer

Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): N.C. D'Amato, N.S. Spoelstra, A. D'Alessandro, K.C. Hansen

Study supervision: J.K. Richer

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# Cancer Research

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## A TDO2-AhR Signaling Axis Facilitates Anoikis Resistance and Metastasis in Triple-Negative Breast Cancer

Nicholas C. D'Amato, Thomas J. Rogers, Michael A. Gordon, et al.

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## **Abstracts:**

### **Endogenous activation of the aryl hydrocarbon receptor promotes triple-negative breast cancer metastasis**

T. Rogers<sup>1</sup>, N. D'Amato<sup>1</sup>, M. Gordon<sup>1</sup>, L. Greene<sup>1</sup>, D. Cochrane<sup>1</sup>, N. Spoelstra<sup>1</sup>, T. Nemkov<sup>2</sup>, A. D'Alessandro<sup>2</sup>, K. Hansen<sup>2</sup>, and J. Richer<sup>1</sup>

<sup>1</sup>Dept. of Pathology, and <sup>2</sup>Dept. of Biochemistry and Molecular Genetics, University of Colorado-Anschutz Medical Campus (Aurora, CO)

**Background:** Resistance to “anoikis” or detachment-induced apoptosis, is critical for metastasis, facilitating survival of carcinoma cells in transit. Anoikis resistance may contribute to the high propensity of triple-negative breast cancer (TNBC) to rapidly metastasize.

**Methods:** Global gene expression was assessed in TNBC cells in attached versus forced-suspension culture. In addition, global metabolomics of intracellular and secreted metabolites produced by attached or suspended cells was performed using UPLC-MS. Aryl hydrocarbon receptor (AhR) activity was measured using a luciferase reporter and qRT-PCR for endogenous AhR target genes.

**Results:** Profiling of TNBC cells in attached versus forced-suspension culture revealed significant upregulation of AhR itself and downstream targets including *CYP1B1*. Recently, activation of AhR via the tryptophan metabolite kynurenine (Kyn) has been found to promote brain tumor growth and motility. Interestingly, genes involved in the Kyn pathway, including the rate limiting enzyme tryptophan-2,3-dioxygenase (*TDO2*), were upregulated in suspended TNBC cells. Metabolite analysis identified a significant increase in Kyn levels in suspended TNBC cells. **Therefore, I proposed that upregulated Kyn-AhR signaling promotes anoikis resistance and metastasis of TNBC.** Indeed, pharmacological inhibition of *TDO2* reduced Kyn production and Kyn-mediated AhR activation in suspended TNBC cells, resulting in reduced survival in suspension. Importantly, *TDO2* inhibition significantly decreased lung colonization and outgrowth *in vivo*.

**Conclusions:** Increased Kyn-AhR signaling may contribute to the TNBC metastasis. Thus, inhibiting *TDO2*-mediated Kyn production may provide a targeted therapeutic strategy to reduce mortality from this aggressive form of breast cancer.

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## Targeting a Kynurenine-Driven Autocrine Loop to Block Triple-Negative Breast Cancer Metastasis

Thomas J. Rogers<sup>1</sup>, Nicholas C. D'Amato<sup>1</sup>, Michael A. Gordon<sup>1</sup>, Lisa I. Greene<sup>1</sup>, Dawn R. Cochrane<sup>1</sup>, Nicole S. Spoelstra<sup>1</sup>, Travis G. Nemkov<sup>2</sup>, Angelo D'Alessandro<sup>2</sup>, Kirk C. Hansen<sup>2</sup>, and Jennifer K. Richer<sup>1</sup>

<sup>1</sup>Department of Pathology, University of Colorado-Anschutz Medical Campus (Aurora, CO),

<sup>2</sup>Department of Biochemistry and Molecular Genetics, University of Colorado-Anschutz Medical Campus (Aurora, CO)

**Background:** Anoikis resistance is a critical trait of metastasis, facilitating survival of carcinoma cells and progression through the metastatic cascade. This ability to resist detachment-induced cell death may contribute to the high propensity for metastasis of triple-negative breast cancer (TNBC), which lacks estrogen and progesterone receptors or HER2 amplification.

**Methods:** TNBC cells in attached versus forced-suspension culture were analyzed for gene expression using Affymetrix Gene Array2.0. Global metabolic profiling of intracellular and secreted metabolites was performed by ultra-performance liquid chromatography tandem mass spectrometry (UPLC-MS). Kynurenine (Kyn) secretion was measured by high-performance liquid chromatography (HPLC). Aryl hydrocarbon receptor (AhR) activity was measured using a luciferase reporter vector and by qRT-PCR for AhR target genes.

**Results:** Gene profiling revealed significant upregulation of multiple genes involved in the kynurenine pathway (KP), including the rate limiting enzymes tryptophan-2,3-dioxygenase (TDO2) and kynureninase (KYNU). Furthermore, metabolic profiling of intracellular and secreted metabolites identified two intermediate metabolites of the KP, formylkynurenine and Kyn, as intracellular metabolites with the *highest* fold increase in suspended TNBC cells. Kyn exhibited the third-highest fold increase among secreted metabolites. Interestingly, AhR, a known Kyn activated transcription factor, was also increased in TNBC cells in suspension. **Therefore, I postulated that upregulation of the KP and subsequent activation of AhR promotes anoikis resistance and consequent invasive capacity of TNBC.** Targeting TDO2 using a pharmacological inhibitor (680C91) reduced both Kyn secretion and Kyn-mediated activation of AhR in suspended TNBC cells. Inhibition or shRNA knockdown of TDO2 or AhR increased anoikis sensitivity and decreased TNBC proliferation and migration/invasion *in vitro*. Furthermore, TDO2 inhibition significantly decreased lung colonization and tumor outgrowth *in vivo* (P<0.001).

**Conclusions:** Increased TDO2 activity may contribute to the metastatic capacity of TNBC. Thus, inhibiting TDO2-mediated tryptophan catabolism may provide a targeted therapeutic strategy to reduce mortality from this aggressive form of breast cancer.

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## **Suppression of CD8 T-cells by tryptophan catabolism in triple-negative breast cancer**

Lisa I. Greene, Tullia C. Bruno, Thomas J. Rogers, Jill E. Slansky, Virginia F. Borges, Jennifer K. Richer

University of Colorado Anschutz Medical Campus, Aurora CO

**Background:** Cancer immune evasion is increasingly recognized as a critical feature of metastatic disease. Conversion of tryptophan (trp) into kynurenine (kyn) and the subsequent secretion of kyn may be a mechanism of immune evasion, given that trp depletion triggers T-cell death, and that kyn binds to the aryl-hydrocarbon receptor (AhR, a ligand-activated transcription factor) in immune cells, often with suppressive consequences. **Methods:** In a screen to identify pathways promoting metastasis, triple-negative breast cancer (TNBC) cells were plated in adherent or forced suspension culture for 24 hours. Expression of the trp catabolizing enzyme trp 2,3-dioxygenase (TDO2) mRNA (qPCR) and protein (immunoblotting) were measured in adherent and suspended cells. Kyn was measured using HPLC. CD8 T-cells were isolated from the blood of human donors and activated via CD3/CD28 stimulation. T-cell death was measured using a viability stain. Flow cytometry was used to measure T-cell markers. **Results:** TDO2 was increased in suspended TNBC cells by at the mRNA level and protein levels. Secreted kyn increased by 2.4-fold in suspended BT549 cells, and this was reversed by a TDO2 inhibitor ( $p < 0.0001$ ). During activation, kyn reduced CD8 T-cell viability ( $n=9$  donors,  $p < 0.05$ ), and these effects were reduced by an AhR antagonist ( $n=6$ ,  $p < 0.05$ ). Consistent with these results, culturing CD8 T-cells in conditioned media (CM) from suspended BT549 cells during activation reduced CD8 T-cell viability ( $n=6$ ,  $p < 0.05$ ) compared to culture in CM from adherent cells. Expression of multiple proinflammatory cytokines by CD8 T-cells cultured in CM from suspended cells was also reduced ( $n=6$ ,  $p < 0.05$ ). **Conclusions:** These results suggest a role for TNBC-mediated trp-to-kyn catabolism in suppressing CD8 T-cells. Further studies will indicate whether TDO2 might be a rational target for improving antitumor immunity in TNBC.

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